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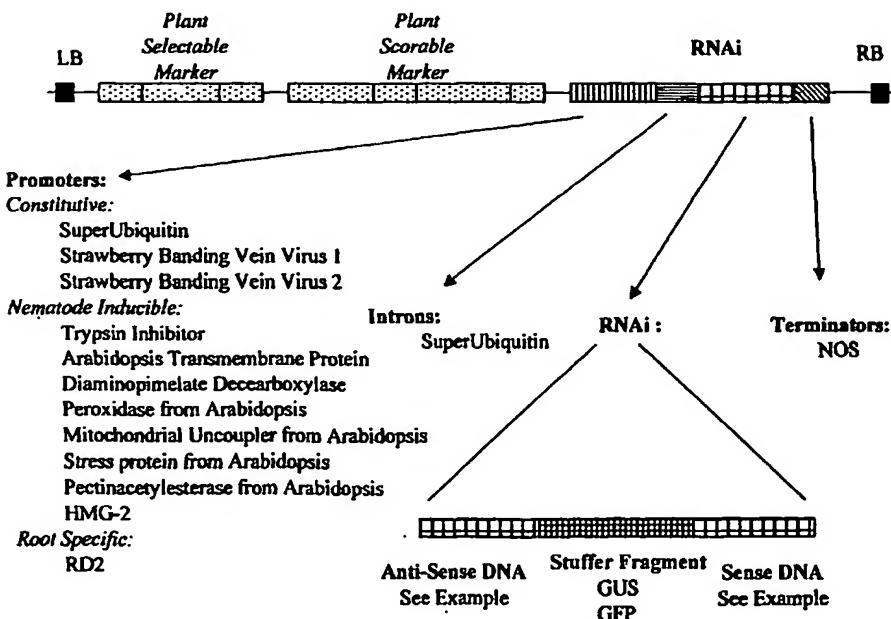
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(54) Title: MATERIALS AND METHODS FOR THE CONTROL OF NEMATODES



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(57) Abstract: The subject invention provides novel methods and compositions for controlling nematodes. More specifically, the subject invention provides RNAi molecules, polynucleotide sequences, and methods of using these sequences in nematode control.



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DESCRIPTIONMATERIALS AND METHODS FOR THE CONTROL OF NEMATODESBackground of the Invention

[0001] Plant parasitic nematodes, such as root-knot nematodes (*Meloidogyne* species) and cyst nematodes (*Globodera* and *Heterodera*), attack nearly every food crop, and are among the world's most damaging agricultural pests. For example, root-knot nematodes parasitize more than 2,000 plant species from diverse plant families and represent a tremendous threat to crop production world-wide. These biotrophic pathogens have evolved highly specialized and complex feeding relationships with their hosts.

[0002] Nematodes cause millions of dollars of damage each year to turf grasses, ornamental plants, and food crops. Efforts to eliminate or minimize damage caused by nematodes in agricultural settings have typically involved the use of soil fumigation with materials such as chloropicrin, methyl bromide, and dazomet, which volatilize to spread the active ingredient throughout the soil. Such fumigation materials can be highly toxic and may create an environmental hazard. Various non-fumigant chemicals have also been used, but these too create serious environmental problems and can be highly toxic to humans.

[0003] Some research articles have been published concerning the effects of δ -endotoxins from *B. thuringiensis* species on the viability of nematodes. See, for example, Bottjer, Bone and Gill ([1985] *Experimental Parasitology* 60:239-244); Ignoffo and Dropkin (Ignoffo, C.M., Dropkin, V.H. [1977] *J. Kans. Entomol. Soc.* 50:394-398); and Ciordia, H. and W.E. Bizzell ([1961] *Jour. of Parasitology* 47:41 [abstract]). Several patents have issued describing the control of nematodes with *B.t.* See, for example, U.S. Patent Nos. 4,948,734; 5,093,120; 5,281,530; 5,426,049; 5,439,881; 5,236,843; 5,322,932; 5,151,363; 5,270,448; 5,350,577; 5,667,993; and 5,670,365. The development of resistance by insects to *B.t.* toxins is one obstacle to the successful use of such toxins.

[0004] The pesticidal activity of avermectins is well known. The avermectins are disaccharide derivatives of pentacyclic, 16-membered lactones. They can be divided into four major compounds: A_{1a}, A_{2a}, B_{1a}, and B_{2a}; and four minor compounds: A_{1b}, A_{2b}, B_{1b}, and B_{2b}. The isolation and purification of these compounds is also described in U.S. Patent No. 4,310,519, issued January 12, 1982. Avermectin B_{2a} is active against the root-knot nematode, *Meloidogyne incognita*. It is reported to be 10-30 times as potent as commercial contact nematicides when incorporated into soil at 0.16-0.25 kg/ha (Boyce Thompson Institute for Plant Research 58th Annual Report [1981]; Putter, I. *et al.* [1981] "Avermectins: Novel Insecticides, Acaracides, and Nematicides from a Soil Microorganism," *Experientia* 37:963-964). Avermectin B_{2a} is not toxic to tomatoes or cucumbers at rates of up to 10 kg/ha.

[0005] Fatty acids are a class of natural compounds which occur abundantly in nature and which have interesting and valuable biological activities. Tarjan and Cheo (Tarjan, A.C., P.C. Cheo [1956] "Nematocidal Value of Some Fatty Acids," Bulletin 332, Contribution 884, Agricultural Experiment Station, University of Rhode Island, Kingston, 41 pp.) report the activity of certain fatty acids against nematodes. In 1977 Sitaramaiah and Singh (Sitaramaiah, K., R.S. Singh [1977] *Indian J. Nematol.* 7:58-65) also examined the response of nematodes to fatty acids. The results of these tests with short chain acids were equivocal, showing nematode-inhibitory action in some instances and stimulatory activity in other instances. Phytotoxicity of these acids was observed at higher concentrations. The short chain fatty acids were also examined by Malik and Jairajpuri (Malik, Z., M.S. Jairajpuri [1977] *Nematol. medit.* 12:73-79), who observed nematode toxicity at high concentrations of the fatty acids.

[0006] Notwithstanding the foregoing (some of the limitations of and problems associated with these approaches are discussed above), there is a need for safe and effective alternatives for controlling nematodes.

[0007] One method for disrupting normal cellular processes is by the use double-stranded interfering RNA (RNAi), or RNA-mediated interference (RNAi). When RNAi corresponding to a sense and antisense sequence of a target mRNA is introduced into a cell, the targeted mRNA is degraded and protein translation of that message is stopped. Although not yet fully understood, the mechanism of this post-transcriptional gene

silencing appears to be at least partially due to the generation of small RNA molecules, about 21 - 25 nucleotides in length, that correspond to the sense and antisense pieces of the RNAi introduced into the cell (Bass, B. L. [2000] "Double-stranded RNA as a template for gene silencing" *Cell* 101:235-238).

[0008] The specificity of this gene silencing mechanism appears to be extremely high, blocking expression only of targeted genes, while leaving other genes unaffected. A recent example of the use of RNAi; to inhibit genetic function in plants used *Agrobacterium tumefaciens*-mediated transformation of *Arabidopsis thaliana* (Chuang, C.-F. and E. M. Meyerowitz [2000] "Specific and heritable genetic interference by double-stranded RNA in *Arabidopsis thaliana*" *Proc. Natl. Acad. Sci. USA* 97:4985-4990). Chuang *et al.* describe the construction of vectors delivering variable levels of RNAi targeted to each of four genes involved in floral development. Severity of abnormal flower development varied between transgenic lines. For one of the genes, AGAMOUS (AG), a strong correlation existed between declining accumulation of mRNA and increasingly severe phenotypes, suggesting that AG-specific endogenous mRNA is the target of RNAi.

Brief Summary of the Invention

[0009] The subject invention provides novel methods and compositions for controlling nematodes. More specifically, the subject invention provides polynucleotide sequences that encode nematode genes, RNAi that selectively targets mRNA transcripts of these essential nematode genes, and methods of using these sequences in nematode control strategies. Such sequences for use according to the subject invention are summarized in Appendix 1. RNAi molecules disclosed herein can be used to inhibit the expression of one or more of these genes in nematodes.

Brief Description of the Drawings

[00010] **Figure 1:** Modular Binary Construct System (MBCS): A series of six, 8-base cutter restriction enzyme sites has been placed between the left and right Ti borders of a previously created kan^R/tet^R binary plasmid.

[00011] **Figure 2:** An exemplary shuttle vector created for cloning of useful DNA fragments by containing the multi-cloning site (MCS) of a modified Bluescript plasmid flanked by 8-base restriction sites.

[00012] **Figure 3:** An exemplary shuttle vector with exemplary inserts.

[00013] **Figure 4:** A suggested RNAi binary vector with exemplary inserts.

[00014] **Figure 5:** Exemplary selectable markers for MBCS.

[00015] **Figure 6:** Exemplary scorable markers for MCBS.

[00016] **Figure 7:** Exemplary RNAi binary vector.

[00017] **Figure 8:** Exemplary RNAi shuttle vector.

Brief Description of the Sequences

[00018] Brief Description of the Sequences can be found in Appendix I.

Detailed Disclosure of the Invention

[00019] The subject invention provides novel methods and compositions for controlling nematodes. More specifically, the subject invention provides polynucleotide sequences and methods of using these sequences in nematode control strategies. A preferred method for controlling nematodes according to the subject invention provides materials and methods for controlling nematodes by using double-stranded interfering RNA (RNAi), or RNA-mediated interference (RNAi). The terms RNAi and RNAi are used interchangeably herein unless otherwise noted.

[00020] In one embodiment of the invention, RNAi molecules are provided which are useful in methods of killing nematodes and/or inhibiting their growth, development, parasitism or reproduction. RNAi molecules of the invention are also useful for the regulation of levels of specific mRNA in nematodes.

[00021] dsRNA (RNAi) typically comprises a polynucleotide sequence identical to a target gene (or fragment thereof) linked directly, or indirectly, to a polynucleotide

sequence complementary to the sequence of the target gene (or fragment thereof). The dsRNA may comprise a polynucleotide linker (stuffer) sequence of sufficient length to allow for the two polynucleotide sequences to fold over and hybridize to each other; however, a linker sequence is not necessary. The linker (stuffer) sequence is designed to separate the antisense and sense strands of RNAi significantly enough to limit the effects of steric hindrances and allow for the formation of dsRNA molecules.

[00022] RNA containing a nucleotide sequence identical to a fragment of the target gene is preferred for inhibition; however, RNA sequences with insertions, deletions, and point mutations relative to the target sequence can also be used for inhibition. Sequence identity may be optimized by sequence comparison and alignment algorithms known in the art (see Gribskov and Devereux, *Sequence Analysis Primer*, Stockton Press, 1991, and references cited therein) and calculating the percent difference between the nucleotide sequences by, for example, the Smith-Waterman algorithm as implemented in the BESTFIT software program using default parameters (e.g., University of Wisconsin Genetic Computing Group). Alternatively, the duplex region of the RNA may be defined functionally as a nucleotide sequence that is capable of hybridizing with a fragment of the target gene transcript.

[00023] As disclosed herein, 100% sequence identity between the RNA and the target gene is not required to practice the present invention. Thus the invention has the advantage of being able to tolerate sequence variations that might be expected due to genetic mutation, strain polymorphism, or evolutionary divergence.

[00024] RNA may be synthesized either *in vivo* or *in vitro*. Endogenous RNA polymerase of the cell may mediate transcription *in vivo*, or cloned RNA polymerase can be used for transcription *in vivo* or *in vitro*. For transcription from a transgene *in vivo* or an expression construct, a regulatory region (e.g., promoter, enhancer, silencer, splice donor and acceptor, polyadenylation) may be used to transcribe the RNA strand (or strands). Inhibition may be targeted by specific transcription in an organ, tissue, or cell type; stimulation of an environmental condition (e.g., infection, stress, temperature, chemical inducers); and/or engineering transcription at a developmental stage or age. The RNA strands may or may not be polyadenylated; the RNA strands may or may not be capable of being translated into a polypeptide by a cell's translational apparatus. RNA

may be chemically or enzymatically synthesized by manual or automated reactions. The RNA may be synthesized by a cellular RNA polymerase or a bacteriophage RNA polymerase (e.g., T3, T7, SP6). The use and production of an expression construct are known in the art (see, for example, WO 97/32016; U.S. Pat. Nos. 5,593,874; 5,698,425; 5,712,135; 5,789,214; and 5,804,693; and the references cited therein). If synthesized chemically or by *in vitro* enzymatic synthesis, the RNA may be purified prior to introduction into the cell. For example, RNA can be purified from a mixture by extraction with a solvent or resin, precipitation, electrophoresis, chromatography, or a combination thereof. Alternatively, the RNA may be used with no or a minimum of purification to avoid losses due to sample processing. The RNA may be dried for storage or dissolved in an aqueous solution. The solution may contain buffers or salts to promote annealing, and/or stabilization of the duplex strands.

[00025] Preferably and most conveniently, RNAi can be targeted to an entire polynucleotide sequence of a gene set forth herein. Preferred RNAi molecules of the instant invention are highly homologous or identical to the polynucleotides summarized in Appendix 1. The homology is preferably greater than 90% and is most preferably greater than 95%.

[00026] Fragments of genes can also be targeted. These fragments are typically in the approximate size range of about 20 nucleotides. Thus, targeted fragments are preferably at least about 15 nucleotides. In certain embodiments, the gene fragment targeted by the RNAi molecule is about 20-25 nucleotides in length. However, other size ranges can also be used. For example, using a *C. elegans* microinjection assay, RNAi "fragments" of about 60 nucleotides with between 95 and 100% identity (to a nematode gene) were determined to cause excellent inhibition.

[00027] Thus, RNAi molecules of the subject invention are not limited to those that are targeted to the full-length polynucleotide or gene. The nematode gene product can be inhibited with a RNAi molecule that is targeted to a portion or fragment of the exemplified polynucleotides; high homology (90-95%) or identity is also preferred, but not necessarily essential, for such applications.

[00028] The polynucleotide sequences identified in Appendix A and shown in the Sequence ID listing are from genes encoding nematode proteins having the functions

shown in Appendix 1. The genes exemplified herein are representative of particular classes of proteins which are preferred targets for disruption according to the subject invention. These classes of proteins include, for example, proteins involved in ribosome assembly; neurol transmitter receptors and ligands; electron transport proteins; metabolic pathway proteins; and protein and polynucleotide production, folding, and processing proteins.

[00029] Genetic regulatory sequences, such as promoters, enhancers, and terminators, can be used in genetic constructs to practice the subject invention. Such constructs themselves can also be used for nematode control. Various constructs can be used to achieve expression in specific plant tissues (by using root specific promoters, for example) and/or to target specific nematode tissues (by using targeting elements or adjacent targeting sequences, for example).

[00030] In a specific embodiment of the subject invention, plant cells, preferably root cells, are genetically modified to produce at least one RNAi that is designed to be taken up by nematodes during feeding to block expression (or the function of) of a target gene. As is known in the art, RNAi can target and reduce (and, in some cases, prevent) the translation of a specific gene product. RNAi can be used to reduce or prevent message translation in any tissue of the nematode because of its ability to cross tissue and cellular boundaries. Thus, RNAi that is contacted with a nematode by soaking, injection, or consumption of a food source will cross tissue and cellular boundaries. RNAi can also be used as an epigenetic factor to prevent the proliferation of subsequent generations of nematodes.

[00031] Nematode polynucleotide sequences disclosed herein demonstrate conserved nucleotide motifs among different nematode genera. Conserved nucleotide motifs strongly suggest that these sequences are associated with viability and/or parasitism and are functionally conserved and expressed in both *Meloidogyne incognita* (root-knot nematode) and *Globodera rostochiensis* and *Globdera pallids* (potato cyst nematodes). The use of these polynucleotides, and RNAi inhibitors thereof, is advantageous because such RNAi can be designed to have broad RNAi specificity and are thus useful for controlling a large number of plant parasitic nematodes *in planta*. Because the genes identified in this disclosure are associated with nematode survival

and/or parasitism, RNAi inhibition of these genes (arising from contacting nematodes with compositions comprising RNAi molecules) prevents and/or reduces parasitic nematode growth, development, and/or parasitism.

[00032] Methods of the subject invention include the transformation of plant cells with genes or polynucleotides of the present invention, which can be used to produce nematode inhibitors or RNAi in the plants. In one embodiment, the transformed plant or plant tissue can express RNAi molecules encoded by the gene or polynucleotide sequence introduced into the plant. Other nematode inhibitors contemplated by the invention include antisense molecules specific to the polynucleotide sequences disclosed herein. The transformation of plants with genetic constructs disclosed herein can be accomplished using techniques well known to those skilled in the art and can involve modification of the gene(s) to optimize expression in the plant to be made resistant to nematode infection and infestation. Furthermore, it is known in the art that many tissues of the transgenic plants (such as the roots) can be targeted for transformation.

[00033] RNA-mediated interference (RNAi) of gene expression. Several aspects of root-knot nematode biology make classical genetic studies difficult with this organism. Since root-knot nematodes reproduce by obligatory mitotic parthenogenesis, the opportunity to perform genetic crosses is not available. Microinjection of RNAi can be used to manipulate gene expression in *C. elegans* (Fire, A., S. Xu, M. K. Montgomery, S. A. Kostas, S. E. Driver, and C. C. Mello. [1998] "Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*" *Nature* 391:806- 811). Microinjecting (into adult nematodes) RNAi can turn off specific genes in progeny worms complementary to the coding region of the genes. Moreover, gene inhibition occurs in progeny when RNAi is injected into the body cavity of the adult, indicating the ability of the RNAi to cross cellular boundaries. This RNAi injection method provides a molecular genetic tool that allows for analysis of gene function in root-knot nematodes.

[00034] RNAi can be taken up by *C. elegans* by simply soaking the nematodes in a solution RNAi. This results in targeted inhibition of gene expression in the nematode (Maeda, I., Y. Kohara, M. Yamamoto and A. Sugimoto [1999] "RNAi screening with a non-redundant cDNA set" International Worm Meeting, Madison, WI, abstract 565). Nematodes fed *E. coli* expressing RNAi also demonstrate targeted and

heritable inhibition of gene expression (Sarkissian, M., H. Tabara and C. C. Mello [1999] "A *mut-6* screen for RNAi deficient mutants" International Worm Meeting, Madison, WI, abstract 741; Timmons, L. and A. Fire [1998] "Specific interference by ingested dsRNA" *Nature* 395:854; WO 99/32619, hereby incorporated by reference in its entirety).

[00035] Accordingly, one aspect of the instant invention is directed to the control of nematodes comprising contacting nematodes with compositions comprising RNAi molecules specific to the nematode genes disclosed herein. The contacting step may include soaking the nematodes in a solution containing RNAi molecules, feeding nematodes RNAi molecules contained in microbes or plant cells upon which the nematode feeds, or injecting nematodes with RNAi. Nematodes can also be "contacted" and controlled by RNAi expressed in plant tissues that would be consumed, ingested, or frequented by nematodes.

[00036] The RNAi molecules provided to the nematodes may be specific to a single gene. A "cocktail" of RNAi molecules specific to various segments of a single gene can also be used. In addition, a "multigene cocktail" of RNAi molecules specific to two or more genes (or segments thereof) may be applied to the nematodes according to the subject invention.

[00037] In addition to RNAi uptake mediated by transgenic plants, nematodes can be directly transformed with RNAi constructs of cDNAs encoding secretory or other essential proteins to reduce expression of the corresponding gene. The transgenic animals can be assayed for inhibition of gene product using immunoassays or for reduced virulence on a host. Progeny of affected worms can also be assayed by similar methods.

[00038] Procedures that can be used for the preparation and injection of RNAi include those detailed by Fire *et al.*, (1998; <ftp://ciw1.ciwemb.edu>). Root-knot nematodes can be routinely monoxenically cultured on *Arabidopsis thaliana* roots growing on Gamborg's B-5/Gelrite® media. This nematode-host pathosystem is ideally suited for these microinjection experiments since limited root galling results in the parasitic stages (late J2 through adult females) developing outside of the root for easy accessibility for injecting. Another advantage is the parthenogenic reproduction of root-knot nematodes, which makes fertilization by males unnecessary for egg production. The RNAi can be injected into the body cavity of parasitic stages of root-knot nematodes

feeding on *A. thaliana* roots using microinjection. Control nematodes can be injected in parallel with only buffer or an unrelated RNAi. Injected nematodes can be monitored for egg production, and the eggs can be collected for the assays described below. Female root-knot nematodes will typically survive and lay more than 250 eggs following 1 μ l injection of buffer.

[00039] Alternatively, methods are available for microinjecting materials directly into the plant root cells upon which nematodes feed: giant cells or syncytial cells (Böckenhoff, A. and F.M.W. Grundler [1994] "Studies on the nutrient uptake by the beet cyst nematode *Heterodera schachtii* by *in situ* microinjection of fluorescent probes into the feeding structures in *Arabidopsis thaliana*" *Parasitology* 109:249-254). This provides an excellent test system to screen RNAi molecules for efficacy by directly inhibiting growth and development of the nematode feeding upon the microinjected plant cell, or by reducing fecundity and the ability of said nematode to generate pathogenic or viable progeny.

[00040] There are a number of strategies that can be followed to assay for RNAi gene interference. Inhibition of gene expression by RNAi inhibits the accumulation of the corresponding secretory protein in the esophageal gland cells of transgenic J2 hatched from the eggs produced by the injected nematodes. In the first assay, polyclonal antibodies to the target gene product can be used in immunolocalization studies (Hussey, R. S. [1989] "Monoclonal antibodies to secretory granules in esophageal glands of *Meloidogyne* species" *J. Nematol.* 21:392-398; Borgonie, G, E. van Driessche, C. D. Link, D. de Waele, and A. Coomans [1994] "Tissue treatment for whole mount internal lectin staining in the nematodes *Caenorhabditis elegans*, *Panagrolaimus superbus* and *Acrobeloides maximus*" *Histochemistry* 101:379-384) to monitor the synthesis of the target protein in the gland cells of progeny of the injected nematodes, or in any other nematode tissue that fails to express the essential targeted gene. Interference of endogenous gene activity by the RNAi eliminates binding of the antibodies to secretory granules in the glands, or any other target tissue, of the transgenic nematodes, and can be monitored by these *in situ* hybridization experiments. Control nematodes injected only with the injection buffer can be processed similar to the RNAi treated nematodes.

[00041] Another assay is designed to determine the effect of the RNAi on reducing the virulence of J2 progeny of the injected females. Egg masses from injected females can be transferred singly to *A. thaliana* plates to assess the ability of the transgenic J2 to infect roots. The J2 hatching from the eggs transferred to the plates can be monitored; after 25 days the number of galls with egg laying females can be recorded. The *A. thaliana* roots can also be stained with acid fuschin to enumerate the number of nematodes in the roots. Egg masses from nematodes injected only with the injection buffer can be handled similarly and used as controls. The treatments can be replicated, and the root infection data can be analyzed statistically. These experiments can be used to assess the importance of the target genes in root-knot nematode's virulence or viability. By staining the J2 progeny of the injected females with the antibodies, it can be determined whether RNAi blocks expression of the targeted gene.

[00042] Additional uses of polynucleotides. The polynucleotide sequences exemplified herein can be used in a variety of ways. These polynucleotides can be used in assays for additional polynucleotides and additional homologous genes, and can be used in tracking the quantitative and temporal expression of parasitism genes in nematodes. These polynucleotides can be cloned into microbes for production and isolation of their gene products. Among the many uses of the isolated gene product is the development of additional inhibitors and modifiers. The protein products of the subject polynucleotides can also be used as diagnostic tools. For example, proteins encoded by the parasitism genes, as identified herein, can be used in large scale screenings for additional peptide inhibitors. The use of peptide phage display screening is one method that can be used in this regard. Thus, the subject invention also provides new biotechnological strategies for managing nematodes under sustainable agricultural conditions.

[00043] Antisense technologies can also be used for phytopathogenic nematode control. Antisense technology can be used to interfere with expression of the disclosed endogenous nematode genes. Antisense technology can also be used to alter the components of plants used as targets by the nematodes. For example, the transformation of a plant with the reverse complement of an endogenous gene encoded by a polynucleotide exemplified herein can result in strand co-suppression and gene silencing

or inhibition of a target involved in the nematode infection process. Thus, the subject invention includes transgenic plants (which are preferably made nematode-resistant in this manner, and other organisms including microbes and phages) comprising RNAi or antisense molecules specific to any of the polynucleotides identified herein.

[00044] Polynucleotide probes. DNA possesses a fundamental property called base complementarity. In nature, DNA ordinarily exists in the form of pairs of anti-parallel strands, the bases on each strand projecting from that strand toward the opposite strand. The base adenine (A) on one strand will always be opposed to the base thymine (T) on the other strand, and the base guanine (G) will be opposed to the base cytosine (C). The bases are held in apposition by their ability to hydrogen bond in this specific way. Though each individual bond is relatively weak, the net effect of many adjacent hydrogen bonded bases, together with base stacking effects, is a stable joining of the two complementary strands. These bonds can be broken by treatments such as high pH or high temperature, and these conditions result in the dissociation, or "denaturation," of the two strands. If the DNA is then placed in conditions which make hydrogen bonding of the bases thermodynamically favorable, the DNA strands will anneal, or "hybridize," and reform the original double- stranded DNA. If carried out under appropriate conditions, this hybridization can be highly specific. That is, only strands with a high degree of base complementarity will be able to form stable double-stranded structures. The relationship of the specificity of hybridization to reaction conditions is well known. Thus, hybridization may be used to test whether two pieces of DNA are complementary in their base sequences. It is this hybridization mechanism which facilitates the use of probes of the subject invention to readily detect and characterize DNA sequences of interest.

[00045] The specifically exemplified polynucleotides of the subject invention can themselves be used as probes. Additional polynucleotide sequences can be added to the ends of (or internally in) the exemplified polynucleotide sequences so that polynucleotides that are longer than the exemplified polynucleotides can also be used as probes. Thus, isolated polynucleotides comprising one or more of the exemplified sequences are within the scope of the subject invention. Polynucleotides that have less nucleotides than the exemplified polynucleotides can also be used and are contemplated within the scope of the present invention. For example, for some purposes, it might be

useful to use a conserved sequence from an exemplified polynucleotide wherein the conserved sequence comprises a portion of an exemplified sequence. Thus, polynucleotides of the subject invention can be used to find additional, homologous (wholly or partially) genes.

[00046] Probes of the subject invention may be composed of DNA, RNA, or PNA (peptide nucleic acid). The probe will normally have at least about 10 bases, more usually at least about 17 bases, and may have about 100 bases or more. Longer probes can readily be utilized, and such probes can be, for example, several kilobases in length. The probe sequence is designed to be at least substantially complementary to a portion of a gene encoding a protein of interest. The probe need not have perfect complementarity to the sequence to which it hybridizes. The probes may be labeled utilizing techniques that are well known to those skilled in this art.

[00047] One approach for the use of the subject invention as probes entails first identifying DNA segments that are homologous with the disclosed nucleotide sequences using, for example, Southern blot analysis of a gene bank. Thus, it is possible, without the aid of biological analysis, to know in advance the probable activity of many new polynucleotides, and of the individual gene products expressed by a given polynucleotide. Such an analysis provides a rapid method for identifying commercially valuable compositions.

[00048] One hybridization procedure useful according to the subject invention typically includes the initial steps of isolating the DNA sample of interest and purifying it chemically. Either lysed nematodes or total fractionated nucleic acid isolated from nematodes can be used. Cells can be treated using known techniques to liberate their DNA (and/or RNA). The DNA sample can be cut into pieces with an appropriate restriction enzyme. The pieces can be separated by size through electrophoresis in a gel, usually agarose or acrylamide. The pieces of interest can be transferred to an immobilizing membrane.

[00049] The particular hybridization technique is not essential to the subject invention. As improvements are made in hybridization techniques, they can be readily applied.

[00050] The probe and sample can then be combined in a hybridization buffer solution and held at an appropriate temperature until annealing occurs. Thereafter, the membrane is washed free of extraneous materials, leaving the sample and bound probe molecules typically detected and quantified by autoradiography and/or liquid scintillation counting. As is well known in the art, if the probe molecule and nucleic acid sample hybridize by forming a strong non-covalent bond between the two molecules, it can be reasonably assumed that the probe and sample are essentially identical or very similar. The probe's detectable label provides a means for determining in a known manner whether hybridization has occurred.

[00051] In the use of the nucleotide segments as probes, the particular probe is labeled with any suitable label known to those skilled in the art, including radioactive and non-radioactive labels. Typical radioactive labels include ^{32}P , ^{35}S , or the like. Non-radioactive labels include, for example, ligands such as biotin or thyroxine, as well as enzymes such as hydrolases or peroxidases, or the various chemiluminescers such as luciferin, or fluorescent compounds like fluorescein and its derivatives. In addition, the probes can be made inherently fluorescent as described in International Application No. WO 93/16094.

[00052] Various degrees of stringency of hybridization can be employed. The more stringent the conditions, the greater the complementarity that is required for duplex formation. Stringency can be controlled by temperature, probe concentration, probe length, ionic strength, time, and the like. Preferably, hybridization is conducted under moderate to high stringency conditions by techniques well known in the art, as described, for example, in Keller, G.H., M.M. Manak (1987) *DNA Probes*, Stockton Press, New York, NY., pp. 169-170.

[00053] As used herein "moderate to high stringency" conditions for hybridization refers to conditions that achieve the same, or about the same, degree of specificity of hybridization as the conditions "as described herein." Examples of moderate to high stringency conditions are provided herein. Specifically, hybridization of immobilized DNA on Southern blots with ^{32}P -labeled gene-specific probes was performed using standard methods (Maniatis *et al.*). In general, hybridization and subsequent washes were carried out under moderate to high stringency conditions that

allowed for detection of target sequences with homology to sequences exemplified herein. For double-stranded DNA gene probes, hybridization was carried out overnight at 20-25° C below the melting temperature (Tm) of the DNA hybrid in 6X SSPE, 5X Denhardt's solution, 0.1% SDS, 0.1 mg/ml denatured DNA. The melting temperature is described by the following formula from Beltz *et al.* (1983):

[00054] $Tm = 81.5^{\circ}\text{C} + 16.6 \cdot \text{Log}[\text{Na}^+] + 0.41(\%G+C) - 0.61(\%\text{formamide}) - 600/\text{length of duplex in base pairs.}$

Washes are typically carried out as follows:

- (1) Twice at room temperature for 15 minutes in 1X SSPE, 0.1% SDS (low stringency wash).
- (2) Once at $Tm - 20^{\circ}\text{C}$ for 15 minutes in 0.2X SSPE, 0.1% SDS (moderate stringency wash).

[00055] For oligonucleotide probes, hybridization was carried out overnight at 10-20° C below the melting temperature (Tm) of the hybrid in 6X SSPE, 5X Denhardt's solution, 0.1% SDS, 0.1 mg/ml denatured DNA. Tm for oligonucleotide probes was determined by the following formula from Suggs *et al.* (1981):

[00056] $Tm (\text{ }^{\circ}\text{C}) = 2(\text{number T/A base pairs}) + 4(\text{number G/C base pairs})$

[00057] Washes were typically carried out as follows:

[00058] (1) Twice at room temperature for 15 minutes 1X SSPE, 0.1% SDS (low stringency wash).

[00059] (2) Once at the hybridization temperature for 15 minutes in 1X SSPE, 0.1% SDS (moderate stringency wash).

[00060] In general, salt and/or temperature can be altered to change stringency. With a labeled DNA fragment of greater than about 70 or so bases in length, the following conditions can be used:

Low: 1 or 2X SSPE, room temperature

Low: 1 or 2X SSPE, 42° C

Moderate: 0.2X or 1X SSPE, 65° C

High: 0.1X SSPE, 65° C.

[00061] Duplex formation and stability depend on substantial complementarity between the two strands of a hybrid, and, as noted above, a certain degree of mismatch

can be tolerated. Therefore, polynucleotide sequences of the subject invention include mutations (both single and multiple), deletions, and insertions in the described sequences, and combinations thereof, wherein said mutations, insertions, and deletions permit formation of stable hybrids with a target polynucleotide of interest. Mutations, insertions, and deletions can be produced in a given polynucleotide sequence using standard methods known in the art. Other methods may become known in the future.

[00062] The mutational, insertional, and deletional variants of the polynucleotide sequences of the invention can be used in the same manner as the exemplified polynucleotide sequences so long as the variants have substantial sequence similarity with the original sequence. As used herein, substantial sequence similarity refers to the extent of nucleotide similarity that is sufficient to enable the variant polynucleotide to function in the same capacity as the original sequence. Preferably, this similarity is greater than 50%; more preferably, this similarity is greater than 75%; and most preferably, this similarity is greater than 90%. The degree of similarity needed for the variant to function in its intended capacity will depend upon the intended use of the sequence. It is well within the skill of a person trained in this art to make mutational, insertional, and deletional mutations that are designed to improve the function of the sequence or otherwise provide a methodological advantage.

[00063] PCR technology. Polymerase Chain Reaction (PCR) is a repetitive, enzymatic, primed synthesis of a nucleic acid sequence. This procedure is well known and commonly used by those skilled in this art (see U.S. Patent Nos. 4,683,195; 4,683,202; and 4,800,159; Saiki *et al.*, 1985). PCR is based on the enzymatic amplification of a DNA fragment of interest that is flanked by two oligonucleotide primers that hybridize to opposite strands of the target sequence. The primers are oriented with the 3' ends pointing towards each other. Repeated cycles of heat denaturation of the template, annealing of the primers to their complementary sequences, and extension of the annealed primers with a DNA polymerase result in the amplification of the segment defined by the 5' ends of the PCR primers. Since the extension product of each primer can serve as a template for the other primer, each cycle essentially doubles the amount of DNA fragment produced in the previous cycle. This results in the exponential accumulation of the specific target fragment, up to several million-fold in a

few hours. By using a thermostable DNA polymerase such as *Taq* polymerase, which is isolated from the thermophilic bacterium *Thermus aquaticus*, the amplification process can be completely automated. Other enzymes that can be used are known to those skilled in the art.

[00064] The polynucleotide sequences of the subject invention (and portions thereof such as conserved regions and portions that serve to distinguish these sequences from previously-known sequences) can be used as, and/or used in the design of, primers for PCR amplification. In performing PCR amplification, a certain degree of mismatch can be tolerated between primer and template. Therefore, mutations, deletions, and insertions (especially additions of nucleotides to the 5' end) of the exemplified polynucleotides can be used in this manner. Mutations, insertions and deletions can be produced in a given primer by methods known to an ordinarily skilled artisan.

[00065] The polynucleotide sequences of the instant invention may be "operably linked" to regulatory sequences such as promoters and enhancers. Nucleic acid is "operably linked" when it is placed into a functional relationship with another nucleic acid sequence. For example, DNA for a presequence or secretory leader is "operably linked" to DNA encoding a polypeptide if it is expressed as a preprotein that participates in the secretion of the polypeptide; a promoter or enhancer is "operably linked" to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site is "operably linked" to a coding sequence if it is positioned so as to facilitate translation. Generally, "operably linked" means that the DNA sequences being linked are contiguous, and, in the case of a secretory leader, contiguous and in reading phase. However, enhancers do not have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, synthetic oligonucleotide adaptors or linkers are used in accordance with conventional practice.

[00066] Polynucleotides and proteins. Polynucleotides of the subject invention can be defined according to several parameters. One characteristic is the biological activity of the protein products as identified herein. The proteins and genes of the subject invention can be further defined by their amino acid and nucleotide sequences. The sequences of the molecules can be defined in terms of homology to certain exemplified sequences as well as in terms of the ability to hybridize with, or be amplified by, certain

exemplified probes and primers. Additional primers and probes can readily be constructed by those skilled in the art such that alternate polynucleotide sequences encoding the same amino acid sequences can be used to identify and/or characterize additional genes. The proteins of the subject invention can also be identified based on their immunoreactivity with certain antibodies.

[00067] The polynucleotides and proteins of the subject invention include portions, fragments, variants, and mutants of the full-length sequences as well as fusions and chimerics, so long as the encoded protein retains the characteristic biological activity of the proteins identified herein. As used herein, the terms "variants" or "variations" of genes refer to nucleotide sequences that encode the same proteins or which encode equivalent proteins having equivalent biological activity. As used herein, the term "equivalent proteins" refers to proteins having the same or essentially the same biological activity as the exemplified proteins.

[00068] It will be apparent to a person skilled in this art that genes within the scope of the subject invention can be identified and obtained through several means. The specific genes exemplified herein may be obtained from root-knot nematodes. Genes, or portions or variants thereof, may also be artificially synthesized by, for example, a gene synthesizer.

[00069] Variations of genes may be readily constructed using standard techniques such as site-directed mutagenesis and other methods of making point mutations and by DNA shuffling, for example. In addition, gene and protein fragments can be made using commercially available exonucleases, endonucleases, and proteases according to standard procedures. For example, enzymes such as *Bal*31 can be used to systematically cut off nucleotides from the ends of genes. In addition, genes that encode fragments may be obtained using a variety of restriction enzymes. Proteases may be used to directly obtain active fragments of these proteins. Of course, molecular techniques for cloning polynucleotides and producing gene constructs of interest are also well known in the art. *In vitro* evaluation techniques, such as MAXYGEN's "Molecular Breeding" can also be applied to practice the subject invention.

[00070] Other molecular techniques can also be applied using the teachings provided herein. For example, antibodies raised against proteins encoded by

polynucleotides disclosed herein can be used to identify and isolate proteins from a mixture of proteins. Specifically, antibodies may be raised to the portions of the proteins that are conserved and most distinct from other proteins. These antibodies can then be used to specifically identify equivalent proteins by immunoprecipitation, enzyme linked immunosorbent assay (ELISA), or Western blotting. Antibodies to proteins encoded by polynucleotides disclosed herein, or to equivalent proteins, can readily be prepared using standard procedures known in the art. The genes that encode these proteins can be obtained from various organisms.

[00071] Because of the redundancy of the genetic code, a variety of different DNA sequences can encode the amino acid sequences encoded by the polynucleotide sequences disclosed herein. It is well within the skill of a person trained in the art to create these alternative DNA sequences encoding proteins having the same, or essentially the same, amino acid sequence. These variant DNA sequences are within the scope of the subject invention. As used herein, reference to "essentially the same" sequence refers to sequences that have amino acid substitutions, deletions, additions, or insertions that do not materially affect biological activity. Fragments retaining the characteristic biological activity are also included in this definition.

[00072] A further method for identifying genes and polynucleotides (and the proteins encoded thereby) of the subject invention is through the use of oligonucleotide probes. Probes provide a rapid method for identifying genes of the subject invention. The nucleotide segments that are used as probes according to the invention can be synthesized using a DNA synthesizer and standard procedures.

[00073] The subject invention comprises variant or equivalent proteins (and nucleotide sequences coding for equivalent proteins or for inhibitors of the genes encoding such proteins) having the same or similar biological activity of inhibitors or proteins encoded by the exemplified polynucleotides. Equivalent proteins will have amino acid similarity with an exemplified protein (or peptide). The amino acid and/or nucleotide identity will typically be greater than 60%. Preferably, the identity will be greater than 75%. More preferably, the identity will be greater than 80%, and even more preferably greater than 90%. Most preferably, the identity will be greater than 95%. RNAi molecules will also have corresponding identities in these preferred ranges. These

identities are as determined using standard alignment techniques for determining amino acid and/or nucleotide identity. The identity/similarity will be highest in critical regions of the protein or gene including those regions that account for biological activity or that are involved in the determination of three-dimensional configuration that is ultimately responsible for the biological activity. In this regard, certain amino acid substitutions are acceptable and can be expected if these substitutions are in regions which are not critical to activity or are conservative amino acid substitutions which do not affect the three-dimensional configuration of the molecule. For example, amino acids may be placed in the following classes: non-polar, uncharged polar, basic, and acidic. Conservative substitutions whereby an amino acid of one class is replaced with another amino acid of the same type fall within the scope of the subject invention so long as the substitution does not materially alter the biological activity of the compound. Below is a list of examples of amino acids belonging to various classes

Class of Amino Acid	Examples of Amino Acids
Nonpolar	Ala, Val, Leu, Ile, Pro, Met, Phe, Trp
Uncharged Polar	Gly, Ser, Thr, Cys, Tyr, Asn, Gln
Acidic	Asp, Glu
Basic	Lys, Arg, His

[00074] In some instances, non-conservative substitutions can also be made. The critical factor is that these substitutions must not detract from the ability to manage nematode-caused diseases.

[00075] An "isolated" or "substantially pure" nucleic acid molecule or polynucleotide is a polynucleotide that is substantially separated from other polynucleotide sequences which naturally accompany a nucleic acid molecule. The term embraces a polynucleotide sequence which was removed from its naturally occurring environment by the hand of man. This includes recombinant or cloned DNA isolates,

chemically synthesized analogues and analogues biologically synthesized by heterologous systems. An "isolated" or "purified" protein, likewise, is a protein removed from its naturally occurring environment.

[00076] Recombinant hosts. The genes, antisense, and RNAi polynucleotides within the scope of the present invention can be introduced into a wide variety of microbial or plant hosts. Plant cells can be transformed (made recombinant) in this manner. Microbes, for example, can also be used in the application of RNAi molecules of the subject invention in view of the fact that microbes are a food source for nematodes

[00077] There are many methods for introducing a heterologous gene or polynucleotide into a host cell or cells under conditions that allow for stable maintenance and expression of the gene or polynucleotide. These methods are well known to those skilled in the art. Synthetic genes, such as, for example, those genes modified to enhance expression in a heterologous host (such as by preferred codon usage or by the use of adjoining, downstream, or upstream enhancers) that are functionally equivalent to the genes (and which encode equivalent proteins) can also be used to transform hosts. Methods for the production of synthetic genes are known in the art.

[00078] Where the gene or polynucleotide of interest is introduced via a suitable vector into a microbial host, and said host is applied to the environment in a living state, certain host microbes are preferred. Certain microorganism hosts are known to occupy the phytosphere, phylloplane, phyllosphere, rhizosphere, and/or rhizoplane of one or more crops of interest. These microorganisms can be selected so as to be capable of successfully competing in the particular environment (crop and other habitats) with the wild-type microorganisms, provide for stable maintenance and expression of the gene expressing a polypeptide of interest, and, desirably, provide for improved protection of the protein/peptide from environmental degradation and inactivation.

[00079] A large number of microorganisms is known to inhabit the phylloplane (the surface of the plant leaves) and/or the rhizosphere (the soil surrounding plant roots) of a wide variety of important crops. These microorganisms include bacteria, algae, and fungi. Of particular interest are microorganisms, such as bacteria, e.g., genera *Pseudomonas*, *Erwinia*, *Serratia*, *Klebsiella*, *Xanthomonas*, *Streptomyces*, *Rhizobium*, *Rhodopseudomonas*, *Methylophilus*, *Agrobacterium*, *Acetobacter*, *Lactobacillus*,

Arthrobacter, Azotobacter, Leuconostoc, and Alcaligenes; fungi, particularly yeast, e.g., genera *Saccharomyces, Cryptococcus, Kluyveromyces, Sporobolomyces, Rhodotorula*, and *Aureobasidium*. Of particular interest are the pigmented microorganisms.

[00080] Methods of the subject invention also include the transformation of plants or plant tissue with genes which encode the RNAi molecules of the present invention. In one embodiment, the transformed plant or plant tissue expresses antisense RNA and/or RNAi. Transformation of cells can be made by those skilled in the art using standard techniques. Materials necessary for these transformations are disclosed herein or are otherwise readily available to the skilled artisan.

[00081] Additional methods and formulations for control of pests. Control of nematode pests using the RNAi molecules of the instant invention can be accomplished by a variety of additional methods that would be apparent to those skilled in the art having the benefit of the subject disclosure. A "cocktail" of two or more RNAi molecules can be used to disrupt one or more of the genes identified herein. The "cocktail" of RNAi molecules may be specific to segments of a single gene or the entire gene. A "multigene cocktail" of RNAi molecules specific to two or more genes (or segments thereof) is also encompassed by the instant invention. In another embodiment of the instant invention, the disclosed RNAi molecules, cocktails, and/or multigene cocktails thereof, may be used in conjunction with other known nematode control agents and methodologies. Such cocktails can be used to combat the development of resistance by nematodes to a certain inhibitor or inhibitors.

[00082] Compositions of the subject invention which comprise RNAi molecules and carriers can be applied, themselves, directly or indirectly, to locations frequented by, or expected to be frequented by, nematodes. Microbial hosts which were transformed with polynucleotides that encode RNAi molecules, express said RNAi molecules, and which colonize roots (e.g., *Pseudomonas, Bacillus*, and other genera) can be applied to the sites of the pest, where they will proliferate and be ingested. The result is control of the pest. Thus, methods of the subject invention include, for example, the application of recombinant microbes to the pests (or their locations). The recombinant microbes may also be transformed with more than one RNAi molecule thereby delivering a "cocktail" of RNAi molecules to the nematode pests. A carrier may be any substance suitable for

delivering the RNAi molecules to the nematode. Acceptable carriers are well known in the art and also are commercially available. For example, such acceptable carriers are described in E.W. Martin's *Remington's Pharmaceutical Science*, Mack Publishing Company, Easton, PA.

[00083] All patents, patent applications, provisional applications, and publications referred to or cited herein are incorporated by reference in their entirety to the extent they are not inconsistent with the explicit teachings of this specification.

[00084] Following are examples that illustrate procedures for practicing the invention. These examples should not be construed as limiting. All percentages are by weight and all solvent mixture proportions are by volume unless otherwise noted.

Example 1—Production of Hairy Roots for RNAi Testing

[00085] A hairy root assay system was developed for testing the anti-nematode activity of RNAi molecules.

[00086] *Agrobacterium rhizogenes*: Several *Agrobacterium rhizogenes* strains produce hairy roots on a variety of plant species. *A. rhizogenes* strains, A4, 15834, 8196 and LBA4404 demonstrate hairy root development on tomato and sugar beet, with A4 being the most efficient. The *A. rhizogenes* strain K599 demonstrated very efficient formation on transgenic soybean hairy roots and was also effective on sugar beet and *Arabidopsis*. However, stain K599 failed to produce hairy roots on tomato tissues possibly due to hyper-virulence.

[00087] Hairy root production: Transgenic hairy roots were identified by stable GUS expression in tomato, sugar beet, soybean and *Arabidopsis*. The construct pAKK1401 (pNOS / NPT-II / tNOS // pSU / GUS / tNOS) was used to produce hairy roots when transformed into *A. rhizogenes* strains A4 or K599. Transgenic roots were identified by GUS expression.

Example 2 – Protocol for Electro-competent *Agrobacterium* and Electroporation

[00088] Electro-competent *Agrobacterium* Protocol:

- [00089] 1. Grow *Agrobacterium* overnight in 5 mls LB + antibiotics at 30°C on shaker (for *Agrobacterium rhizogenes* strain K599 no antibiotics are needed).
- [00090] 2. Use the 5 mls of overnight culture to inoculate 500 mls LB + antibiotics at 30°C on shaker. Grow overnight.
- [00091] 3. Add liquid culture in eight 50 ml polypropylene orange cap tubes.
- [00092] 4. Centrifuge 10 min., 4000 rpm, 4°C.
- [00093] 5. Resuspend cells in each tube with 20 mls 10% glycerol (on ice)
- [00094] 6. Centrifuge 10 min., 4000 rpm, 4°C.
- [00095] 7. Resuspend cells in each tube with 10 mls 10% glycerol (on ice).
- [00096] 8. Centrifuge 10 min., 4000 rpm, 4°C.
- [00097] 9. Resuspend cells in each tube with 2 mls 10% glycerol (on ice).
- [00098] 10. Aliquot 50 µl into cold Eppendorf tube and place onto dry ice.
- [00099] 11. Store electro-competent cells at -80°C. These cells can be used for up to two years.

[000100] Electroporations:

- [000101] 1. Add 1 µl to 5 µl of DNA (resuspended in H₂O and not TE or other buffer) to 50 µl of *Agrobacterium* electrocompetent cells and mix.
- [000102] 2. Transfer 20 µl of DNA/*Agrobacterium* mix to cuvette.
- [000103] 3. Electroporate:
25µF, 400 Ω resistance, 2.5 volts (0.2cm cuvette) or 1.8 volts (0.1cm cuvette for BioRad electroporator. 330 µF, 4000 kΩ, low w, fast charge rate for BRL Electroporator.
- [000104] 4. Add 1ml of LB and transfer to Eppendorf tube.
- [000105] 5. Shake at 30°C for 2 hours.
- [000106] 6. Centrifuge down cells (2 min. 14 krpm).
- [000107] 7. Plate all onto LB + antibiotics (most *Agrobacterium* strains are naturally streptomycin resistant).

Example 3 – Protocol for Production of Transgenic Hairy Roots on Soybean

[000108] Seed Sterilization. Rinse the soybean seed with 70% ETOH for 2-5 min. Remove and add 20% Clorox and shake for 20-25 min. Rinse 3X with sterile water. Plate the seed, 5 seed per plate, onto $\frac{1}{2}$ MSB5 + 2% sucrose + 0.2% gel (referred to as $\frac{1}{2}$ MSB5). Place seed into chamber at 25C, 16/8 photoperiod for 5-7 day (depending on genotype) germination period. After 1 week seedlings can be placed into cold room for longer storage if necessary (not to exceed 2 weeks).

[000109] Agrobacterium Preparation. For Agrobacterium rhizogenes strain K599, take a small sample from frozen glycerol into 25-50 ml of NZYM media with 50 mg/L kanamycin in a 125-250 ml Erlenmyer flask. Place onto shaker at 28-30 °C for 16 - 20 hours. Pour sample into centrifuge tube and centrifuge the bacterium at 4000 rpm for 10 min. Pour off supernatant and re-suspend the pellet with an equal volume of liquid $\frac{1}{2}$ MSB5 + 200 μ M acetosyringone. Use pipette to re-suspend the pellet and homogenize the sample (remove all clumps). To determine O.D., prepare a 1:10 dilution by putting 900 μ l $\frac{1}{2}$ MSB5 into cuvette and add 100 μ l of bacterial sample. Determine the O.D.₆₆₀ and calculate the volume needed to adjust (dilute) OD to approximately 0.2 for inoculation. Check final O.D.

[000110] Explant Preparation and inoculation. Place a sterile filter paper onto plates of 1/2 MSB5. Cut soybean cotyledons just above the shoot apex and place onto plate. Lightly scar the cotyledon's abaxial surface (flat side, upper surface that reaches toward sun) with a scalpel blade. Cut each cotyledon transversely into 2-3 pieces (no smaller than 1 cm). Add approximately 10 ml of prepared bacterial solution to each plate and allow cotyledons to incubate for 1 hr. Remove the bacteria using a vacuum aspirator fitted with sterile pipette tip, ensure that there is no standing liquid. Orient all explants with abaxial surface up and wrap plates for a 3 day co-culture, 25°C in light (16/8 photoperiod).

[000111] Hairy root selection and maintenance. After 3 day co-culture, wash explants with liquid $\frac{1}{2}$ MSB5 + 500 mg/L carbenicillin. Transfer the explants abaxial side up to selection media, $\frac{1}{2}$ MSB5 supplemented with 500 mg/L carbenicillin and 200 mg/L kanamycin. Roots should develop in approximately 2-3 weeks. The roots will form primarily from the cut vascular bundles with other roots developing from the small cuts on cotyledon surface. Remove roots (>1cm in length) and place onto replica media with

transfers to fresh media every 2 weeks to prevent *Agrobacterium* overgrowth. After 6-8 weeks on selection the roots can be moved to media without kanamycin, however carbenicillin must remain in media for several months for continued suppression of *Agrobacterium*. At this stage roots can be used for testing RNAi for nematode control. Sterilized nematodes can be added and observed for RNAi affects.

Example 4 – Testing of RNAi for Plant Parasitic Nematode Control.

[000112] Various types of nematodes can be used in appropriate bioassays. For example, *Caenorhabditis elegans*, a bacterial feeding nematode, and plant parasitic nematodes can be used for bioassay purposes. Examples of plant parasitic nematodes include a migratory endo-parasite, *Pratylenchus scribneri* (lesion), and two sedentary endo-parasites, *Meloidogyne javanica* (root-knot) and *Heterodera schachtii* (cyst).

[000113] *C. elegans*: RNAi vectors can be tested through expression of the RNAi in *E. coli*. *C. elegans* are fed *E. coli* and assayed for their growth by measuring growth of nematodes, production of eggs and viability of offspring. Another approach is to inject dsRNA directly into living nematodes. Finally, soaking nematodes in a solution of *in vitro*-prepared RNAi can quickly establish efficacy of treatment.

[000114] *P. scribneri*: The *P. scribneri* *in vitro* feeding assay uses a corn root exudate (CRE) as a feeding stimulus and both the red dye Amaranth or potassium, arsenate as feeding indicators. Feeding is confirmed after seven days by the presence of red stained intestinal cells in live worms exposed to the Amaranth or death of worms exposed to arsenate. This bioassay is used to test soluble toxins or RNAi. *P. scribneri* has also been cultured on wild type roots of corn, rice and *Arabidopsis*, and on *A. rhizogenes*-induced hairy roots of sugar beet and tomato. *P. scribneri* is very valuable in evaluating transgenic hairy roots because of the non-specific feeding of these worms.

[000115] *M. javanica*: Nematode eggs are sterilized using bleach and are used to inoculate hairy roots expressing RNAi. Nematodes are assessed for their growth by measuring knots, egg masses or production of viable eggs. An alternative approach is to microinject dsRNA directly into root feeding sites or into living female nematodes.

[000116] *H. schachtii*: Cultures of this nematode were maintained on sugar beets. Nematodes eggs are sterilized using bleach and used to inoculate hairy roots

expressing RNAi. Nematodes can be assessed for their growth by measuring knots, egg masses or production of viable eggs.

Example 5 – Plant Expression Vectors for RNAi

[000117] Modular Binary Construct System (MBCS): An important aspect of the subject disclosure is the Modular Binary Construct System. The MBCS eases the burden of construct development by creating modular pieces of DNA that can be easily added, removed, or replaced with the use of low frequency cutting restriction enzymes (8-base cutters). These constructs are useful for delivery of a variety of genes to plant cells and is not limited to the delivery of RNAi genes. To develop this system, a series of six, 8-base cutter restriction enzyme sites was placed between the left and right Ti borders of a previously created kan^R/tet^R binary plasmid (Figure 1). The production of both kan^R and tet^R MCBS aids the testing of constructs using different strains of *Agrobacterium rhizogenes* in different plant species. In addition to the MBCS, a series of shuttle vectors were created that aid in the cloning of useful DNA fragments by containing the multi-cloning site (MCS) of a modified Bluescript plasmid flanked by 8-base restriction sites (Figure 2). With six 8-base cutter sites, each site is, preferably, reserved for a particular function (Figures 3 and 4). Because of the close proximity of the *Pme* I and *Sgf*I sites to the left and right border of the binary vector, these sites are, preferably, reserved for gene tagging and enhancer trap experiments. The *Not* I site is, preferably, reserved for plant selectable markers (Figure 5). The *Pac* I site is reserved, preferably, for Plant Scorable Markers (Figure 6). The *Asc* I site is, preferably, reserved for RNAi experiments (Figures 7 and 8), while the *Sbf*I site is, preferably, reserved for anti-nematode proteins. The restriction sites that are denoted in the Figures are, preferably, reserved for the denoted insertions; however, the MCBS binary and shuttle vectors do not require the restriction sites to contain these suggested inserts.

[000118] Plant Selectable Markers for MBCS: To further develop the MBCS, a series of plant selectable markers were added to the MBCS (Figure 5). Plant selectable markers that were added to the MBCS include: pNOS/NPT-II/tNOS (kan^R), pNOS/Bar/tNOS (basta^R for dicots), pUBI/Intron-Bar/tNOS (basta^R for monocots), and pUBI/Intron-PMI/tNOS (mannitol isomerase^R).

[000119] Reporter Genes for MBCS: Four exemplary reporter genes are used in the MBCS are provided in Figure 6 and Appendix 2. GUS, a nuclear localized GUS, GEP, and the anthocyanin transcriptional activator *papIC* genes into the MBCS.

[000120] Promoters for MBCS: We cloned several useful constitutive and nematode-inducible promoters (Figures 6, 7 and Appendix 2). Constitutive promoters include the SuperUbiquitin promoter from pine (pSU) and two promoter regions from the Strawberry Banding Vein virus (pSBV₁ and pSBV₂). Seven nematode-inducible promoters from *Arabidopsis* were also been cloned.

[000121] The following Scorable marker clones have been constructed and placed in the MBCS, NPT-II binary vector (pNOS/NPT-II/tNOS):

Intron/GUS/tNos	Intron/NLS-GUS/tNOS	Intron/GFP/tNOS
pSU/Intron/GUS/tNOS	pSU/Intron/NLS-GUS/tNOS	pSU/Intron/GFP/tNOS
pSBV ₁ /Intron/GUS/tNOS	pSBV ₁ /Intron/NLS-GUS/tNOS	pSBV ₁ /Intron/GFP/tNOS
pSBV ₂ /Intron/GUS/tNOS	pSBV ₂ /Intron/NLS-GUS/tNOS	pSBV ₂ /Intron/GFP/tNOS
pKT/Intron/GFP/tNOS		
pKA/Intron/GFP/tNOS		

Example 6 – Control of Plant parasitic nematodes using RNAi in planta

[000122] Production of RNAi Vector. The RNAi shuttle vector to be used is adapted from the Modular Binary Construct System (MBCS - See Example 5). RNAi shuttle vectors preferably comprise a promoter, intron, antisense RNAi, stuffer fragment, sense RNAi, and terminator (See Figures 7 and 8 and Appendix 2 for more details). The plant promoter can be constitutive, tissue-specific or nematode-inducible. The intron is necessary to eliminate expression in *Agrobacterium*.

[000123] The anti-sense and sense RNAi molecules comprise nematode-specific sequences and are disclosed herein. These genes are associated with pathogenesis, growth, or other cellular function in nematodes. An exemplary group of RNAi sequences for use in plant/nematode control may be based upon:

[000124] 1. Genes specific for nematode esophageal gland cells.

[000125] 2. Genes specific for plant parasitic nematodes but not other free living nematodes.

- [000126] 3. Genes common to all plant parasitic nematodes.
- [000127] 4. Genes common to all nematodes (nematode-specific).
- [000128] 5. Genes specific for important tissues or cell types.
- [000129] 6. Genes from large gene families.
- [000130] 7. Genes involved in nematode signal transduction or other cellular pathways.

[000131] Appropriate RNAi constructs allow for the formation of dsRNA molecules (the sense and antisense strands join to form the dsRNA). The terminator sequence adds a poly-A tail for transcriptional termination. The RNAi shuttle vector can then be subcloned into the MBCS and transformed into *Agrobacterium rhizogenes*.

[000132] Plant Transformation with RNAi Vectors. An exemplary transformation system for generating hairy roots using *Agrobacterium rhizogenes* is provided below. The RNAi vector once introduced into the MBCS can subsequently (as a binary vector) be transformed in *A. rhizogenes* using, for example, the electroporation protocol of Example 2. Once the *A. rhizogenes* is confirmed to contain the plasmid, it is then used in generating hairy roots (See Example 3). Using this protocol transgenic hairy roots expressing RNAi are isolated, cultured and tested.

[000133] Testing of RNAi Vector for Nematode or Plant Pathogen Resistance. RNAi expressing hairy roots can be inoculated with sterilized nematodes. Infested hairy roots can be observed and the effect on nematodes determined. An alternative approach involves the microinjection of RNAi directly into root feeding sites (giant-cells for root-knot nematode, and syncytia for cyst nematodes) or into living female nematodes.

Example 7 – Insertion of Genes Into Plants

[000134] One aspect of the subject invention is the transformation of plants with genes encoding proteins of the present invention. Transformation of plants as described herein can be used to improve the resistance of these plants to attack by the target pest.

[000135] Genes, polynucleotides, and/or RNAi molecules as disclosed or suggested herein can be inserted into plant cells using a variety of techniques which are

well known in the art. For example, a large number of cloning vectors, for example, pBR322, pUC series, M13mp series, pACYC184, pMON, *etc.*, are available for preparation for the insertion of foreign genes into higher plants via injection, biolistics (microparticle bombardment), *Agrobacterium tumefaciens*, or *Agrobacterium rhizogenes*-mediated transformation, or electroporation as well as other possible methods. Once the inserted DNA has been integrated into the genome, the genetically modified-cell(s) can be screened via a vector carried-selectable marker that confers on the transformed plant cells resistance to a biocide or an antibiotic, such as kanamycin, G418, bleomycin, hygromycin, chloramphenicol, or bialaphos, *inter alia*. The transformed cell will be regenerated into a morphologically normal plant. The transgene(s) in the transgenic plant is relatively stable and can be inherited by progeny plants.

[000136] If a transformation event involves a germ line cell, then the inserted DNA an corresponding phenotypic trait(s) will be transmitted to progeny plants. Such plants can be grown in the normal manner and crossed with plants that have the same transformed hereditary factors or other hereditary factors. The resulting hybrid individuals have the corresponding phenotypic properties.

[000137] It should be understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application.

We claim:

1. An RNAi molecule, optionally comprising a linker, wherein at least one strand of said RNAi is encoded by a DNA sequence selected from the group consisting of SEQ ID NO: 1 through SEQ ID NO: 139.
2. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 1.
3. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 2.
4. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 3.
5. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 4.
6. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 5.
7. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 6.
8. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 7.
9. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 8.
10. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 9.

11. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
10.
12. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
11.
13. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
12.
14. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
13.
15. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
14.
16. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
15.
17. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
16.
18. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
17.
19. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
18.
20. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
19.
21. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
20.

22. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
21.

23. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
22.

24. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
23.

25. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
24.

26. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
25.

27. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
26.

28. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
27.

29. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
28.

30. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
29.

31. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
30.

32. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
31.

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33. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
32.

34. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
33.

35. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
34.

36. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
35.

37. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
36.

38. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
37.

39. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
38.

40. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
39.

41. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
40.

42. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
41.

43. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
42.

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44. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
43.

45. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
44.

46. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
45.

47. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
46.

48. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
47.

49. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
48.

50. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
49.

51. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
50.

52. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
51.

53. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
52.

54. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO:
53.

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55. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 54.
56. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 55.
57. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 56.
58. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 57.
59. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 58.
60. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 59.
61. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 60.
62. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 61.
63. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 62.
64. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 63.
65. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 64.

66. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 65.
67. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 66.
68. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 67.
69. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 68.
70. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 69.
71. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 70.
72. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 71.
73. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 72.
74. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 73.
75. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 74.
76. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 75.

77. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 76.
78. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 77.
79. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 78.
80. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 79.
81. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 80.
82. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 81.
83. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 82.
84. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 83.
85. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 84.
86. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 85.
87. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 86.

88. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 87.

89. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 88.

90. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 89.

91. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 90.

92. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 91.

93. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 92.

94. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 93.

95. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 94.

96. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 95.

97. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 96.

98. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 97.

98.

100. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 99.

101. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 100.

102. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 101.

103. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 102.

104. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 103.

105. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 104.

106. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 105.

107. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 106.

108. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 107.

109. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 108.

110. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 109.

111. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 110.

112. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 111.

113. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 112.

114. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 113.

115. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 114.

116. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 115.

117. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 116.

118. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 117.

119. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 118.

120. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 119.

121. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 120.

122. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 121.

123. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 122.

124. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 123.

125. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 124.

126. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 125.

127. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 126.

128. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 127.

129. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 128.

130. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 129.

131. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 130.

132. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 131.

133. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 132.

134. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 133.

135. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 134.

136. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 135.

137. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 136.

138. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 137.

139. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 138.

140. An RNAi molecule according to claim 1, wherein said DNA sequence is SEQ ID NO: 139.

141. A transgenic plant or transgenic plant tissue comprising an RNAi molecule according to any of the preceding claims.

142. A method of disrupting cellular processes in a nematode comprising the steps of:

- (a) providing a composition comprising a compound according to any of the preceding claims; and
- (b) contacting a nematode with said composition.

143. An isolated promoter comprising the following nucleotide sequence:

aacagcccaagataaca~~g~~aaaagtcaaagggtttcgaaa
gaccactt~~t~~tgactaaggatcattcatccataattatctggtagca
cagactcatgataactgcgaggaacacaagttctttacagtgcattc
aaagacactttctttacggttcattgaaggagccgaccagaat
atgcagagaagctttcactgtgggtaatttcattaatctatcca
ggtaaaaacctcaaggagatctctttccaaaagacctctacag
ggcaatcaaaaactacagaaccagagttgttagtgcacagagtagac
caatctacctgagaatcacgagttcttccatagagtggaaaatgat
gacatccttattccataccactggattgaggttaggactatccatgg
aaaaattccatggacaagtcatataagaagaccacaacagtgcagt
atcttccagagataactgcactcagacctaaaggataaaagcagta
tataatcagtgtactaagatctcgcagattcaaagaagacttaa
ctatgctgatgacaagataattctaataagcaattattcagaattaa
tcaaggagaaaagaattataactttcagaatatgaagccgctt
acaagtggccagctagctactgaaaagacagcaagacaatggtg
tctcgatgcaccagaaccacatcttgcagcagatgtgaagcagcca
gagtggtccacaagacgcactcagaaaaggcatcttctaccgacaca
gaaaaagacaaccacagctcatccaaatgttagactgtcgttat
gcgtcggctgaagataagactgaccccaggccagcactaaagaagaa
ataatgcaagtggccatgcacttttagcttataattatgttt
cattattattctctgtttgcctctatataaaagagcttgcatttt
cattgaaggcagaggcgaacacacacacagaacccctgcattaca
aaccatgtattgttagctaaacccctttaggag.

45
144. An isolated promoter comprising the following nucleotide sequence:

tggtggggacaatggatccggtctgcgtacaacaaggctg
aaaaagattaaacagaaacacctgtgatcattagcgtggaccacc
aaaacctcctgagccaccaaagcctccagagcctgaaaaaccaaagc
ctccaccagcacctgaaccaccaaagcatgtatgcacccaccc
tgcaacagttgtgatgtgtctgttactacctatgaaagtggaaag
cggtgcaccattttgagtcataatgcgtaccatagccttcat
gttaagtccctgtatccaaatactaattcatcatgttctcatgct
ttttgtttatccctttctcaaatatgaaatctctgttgc
ctccctgttataattagtcgttgcacacaagaagtctcatg
agttcatgctaaagaaataaaagttcaaaattaaaacaccaaatt
tgattaaattccataaaacacctgtgaaagcagaaagtttagtc
ctgaacagagcttaggaagtcgttgcaggacatatctcaagtgc
ttgggtcgtagcactcttaggcccattacttcattgagcccatt
attatgcaaaacaagaaatgagacatatgaaacattagggttctt
cagaaaaaaataggaaaaagcagggacaactaaacaaaaattcagaa
acaagaggcaagtggacgaccacggcgtaaagatcaacatgtgg
gtgcattgagaccaagaccattttctcggttcaacgcacactt
gtctttcttatgtttgtgcattcttattaggcagaccctct
cttttaataggatagaaaaatatgattttatccatgtgaaa
catttgagttaaaacctaaacttatgatcattgttagt
ttccatagcatacgttatcaacatgacctctaaccaaaaatt
gatgaaactactttaatgatgatcattttatccat
ttaaatttagtagttgtgaaattatgacatgattgcgtc
aatcaaaacagttatcgtgaaacttaggagaatgtttatcgt
gttcaacacatgattgcataatgttaggtgtc
cataacaatcatcactcgtaaatatcaaagtggttctgag
aaagggttatgatttccaaactgcacttagttgttatt
cacacgtatgcattctgagttctgcccagtgaaattaaagc
ttgggagagatcataattattagggttgcattgc
cgtaaaatgaaaatttgcattcttccaaacacaaaagaaat
ctagttatcttttttatataacaattcatgaaatgt
tttatacacatcatccaaatcgaattgtaatct
cagtagatggcaataagatatacaatccatggacc
atgtgtaattcatacatgcattttgtctgcatttatt
atgctgagcgttttaagtgtgaaactagatct
aagatggtctttctgtttgtcgataagagcaag
gatcaattttaaaattctaaataaaactccaaacc
catgaaactcttttagaaaatccatttataaca
aataattc
tgatttttttttttttttttttttttttttttttttttt
ctcagaaaaagccatttttttctattcttgc
tactgtgcgttttacaaagttgtc
actcacagtcacagagatctgttttttttttttt
ttcttccagt.

145. An isolated promoter comprising the following nucleotide sequence:

.agcaaagcaagaacaccagagaagaagaaaagcactacaga
gaaaaatgtgagcttaagcgctctccaaacaacacttctctggagtc
taaaggatgtgcaaaaagccttgggttagacttccgcatttc
caagcatgggttattttgttagcacacaaactatctgaccctcga
cttggatttcttcgcagttgtccaaactacattgaaacggatatg
caggcaacatggatcatgagggtggcatctcgtaagattaacaaag
tgaacaggtcactaaggaaaatacagacggtaactggactcggtccaa
ggtgtagaaggaggactaaagttcgactcagcaactggcgaattcat
tgcagttagacctttattcaagaattgataccaaaagggtctgt
cgtctttgataatgatgcacatgcaagaagaagttaggaggatatg
cctgacgatacttcattcaagctccaggaagctaaatctgtcgaccaa
tgccattaagtttagaggaggatacaccatgaatcaagcaagaccag
gtaagaacttctctatccataaaccatagatggagcgattagaatct
taatccatttcagtttgcaggatcattcatggaggttaatgcta
gtggcagccatgggttggatggccaaagagtctggcttgaatggc
agtgaaggaataaagagcgttgcacttaagctctgtggaaatttc
agatggaatggatccaacaatccgatgcagtggcagtattttgaac
ctaaccatccatgtcatgcagcatacatcagattcatcaaattggctca
ggcgcagttctgcgttggaaagctcatctacttccatggaaagattggaa
ccaaatgagaacccacaacagtaatagcagcagagtggatcaacaa
cgctgatcgtaaaggccagttatagagaagacactgtacgttcaag
ttcgagccatcagttgggttcctcagctctacaaagaagttggaaa
acgttttaactgcaggacgggtcggttcagctgaagttacttggatg
atgaagaagaatgggtgatgctggttacagattctgatctccaaagaa
tgtttggagatattacatggatggaaaacactcggtgaagttct
cgttcgtgattgtctgcccctctaggttagttctggcggcagtaatg
gttatcttggaaacaggcttatgacgtcgtaagacatagacacacaca
gttatgtattcccagtgaagaatgtttttttctctagatatta
gtatgcttataataggcatgaaggagaagacaattttgtatagt
ggagttcagcagaaaatgtatgttttcgttttatgatcag
agaataaaagtggatgttatctacgttgcataatgttacctgc
tcacccatttcatataagaaaagagaacacttttagttatccctg
tgatgcagaatcgatttttttatctccattccgtggaaacc
aacaagtcactaaattcggttaattgggtggtttaagtcaa
cgaggactgttttagttgggttggcctataattgtgttcatca
ttgggtttttcccttatcagtttaacgtccatatccatatctt
ttcttttttaacggcaaagttcatatccatatctttagatgtgcct
aaaagaggagaagatgcgaagacagaatttcatattgaaagggt
tcgatatcgatattggaaacgaatcaaggtcaaaaaactcagtcta
atagttgaatttaaaaattttattatcaatccgattggttcgt
tttggatgttgcgttctatcatcaaaccaatcggttggcct
aaagataattataatattcacaacaccagtgttaaacacatatca
acaaacctaagttagataaacaaagaga.

47
146. An isolated promoter comprising the following nucleotide sequence:

48
147. An isolated promoter comprising the following nucleotide sequence:

48

ttggccaaactgagatataagagggaaagggtgatttcatgcaa
atttttttttatTTTTGAATGAATGCAAAATTATTCAAAA
aaaaaaacTGGCTACATCAAGTACTTCATTCTGAGTTGAAA
aatCTAAAGACAACAAAAGACTTACAATTAAATAAAAAATAAA
AAATACATTATCCTCAACGAAATTGTTGATTAATAACGTATCT
CTTGGTAAAACAGCGTTATTGACGAAATTGTTATAATGAATAA
AATGATAATAGAAAAGCTAGTGTGGTACGTAAAATACCTCTCATTGGC
AAAATAACCGTTATGTATCATGAGTATTGACGACAGCGTGCCTA
AATAGTGTGCTTCAGGAGAAAATATACCAAGTTATTGCTGAAA
TTACCAACGCAAATCTGAGGTTGCAATGGCAAAATAAAAAACCAATGT
CATTTCCTTAATGTATTAAGGTCAATTAAATAAAATTGTACACTTT
TTCACCTGTAAGCGTCCAAAGTGTAGAATGGATAACTAGAAGGGTC
AAAGGTATAATATTAAATAAGCGAACTCACTTTGCCAAGTGTATT
CACTTCTTACATTGCTTGTATAGTTACCCAAAAGTGTATATATAT
TCCCTTATAACAATTGTTCTATTCTGGATTATAAGGGAAATAAGAA
AAAAGAAAAGAGAGAGTATATAATAACTTTATAAAGTGTATGTTA
GATTCTAATTGTAACGAAAAGTTCAAAGTGTAAAGAAAACGAAAAA
AGTTTTCTGTTGTTGTTATATCTATAGCCAAGAAAAGTTCTCAGA
TTTACAAGAAGTTAACTGAGAAAACAAAAAAACTTATGAAGCA
TGAAAGACTAATTACGAGGTGATTAATTGAGACAAATTAAACAT
CGAATTAAGATAACATTGGAGGGTTATATGTTATATGTGACA
TGATAAGTCCGATTCTGACTAATGTATATCTGGAATCTAACATGGA
AGAATAGAGAACGAAAGCAGAGCCAAGGTCAACTTGGCCAGACACGAAT
CAACAGATTGTGAATGAGACCAATCAATGGTCATAAACCGGTTGGG
TTAAACCGGCAAGTCATCCTGGCTCAATTCCATTGTTATTCTT
CATGCAAGACCCCTCTGATACACCAAGACTCCCATTACAATATTCT
TTCGATCACGAGCTACTTATTCAAATGTGTTACCTCTTCGTGAC
TCTTGTGTTGTGGTAAAGCCTAGTCGAGATGTGTCGGTATATATA
GGCATAACATACAAATGCGACAAAATAAGTATATTATATTGTTAA
TTTCTATATTCCATTCTATATGCAATGGCTGGGATTTGACCAAAA
CCCTAATTCAAGAATAGAATCCAAAAGATGGGATCAAAGAATATAAT
CTAATGGGCTGACCACATTCCGATTACATCGCATAGTTAATT
CTTCCACTACTTATGCCGAGAAATTGTAATTAGTAAGACAAA
GAAATACAGATATAAGATGGCTGTAGAAACCAGTAGAGGAATTCT
TTTCGTGGATAAGTGGATATTAAAGAGAATGGCTTTACTCTT
TACAGTGGAAATGGGATAGTAGCCATTATAATTCTCATCAGATT
TATATATGCACTGTTGTATAAGCTAAAATAACGTTAAGCATT
TTCAAAAAAATTACAAGTTCTAGAGACTCTTAAACGTCGGCAATT
TATATTCTACTTTACATGACACTTCAGGAAAAGAAAACATAACTCA
CTAGCAGATCATAAATTCTTTCTTTGAAATGAACCTAG
TTGTGGTTTATTCTGTTAGCTAGAAACTCAGTGTGTTTCC
GCCAATGGTAGTQCTTGTATGATGGTCCGG.

148. An isolated promoter comprising the following nucleotide sequence:

caatcaaggtaacgaaggaggatcagcgaaaggatgggcta
ta~~t~~ttggagttttcctgcgtgtaa~~g~~taatgctt~~g~~tatcttcca
tgcggacatataactgaagaataaactcaactcattgttctggg
t~~t~~tttctctgatcagattc~~t~~cgttgc~~t~~actc~~t~~acttctgt
ggggc~~t~~tattataaaacaagagttagcgtgtggtaatcttcat
atcttctacaattccacttccattcttaattattctcacgtga
tatacacacactcaatcactgtat~~c~~gtatggatc~~g~~agcgtgga
actgatgcattgc~~c~~gggatgtcacttctatc~~g~~ggctactagaaac
tgta~~g~~tattacaagaaaactcaaaggattccatttatgcaaaatc
taagagaaaagctcactgtgtcttgg~~t~~acaatttatggatctc
aagagacaatgtatgtaa~~g~~cttaattgatttgg~~t~~cttgataaaca
gg~~t~~gagtg~~g~~gaa~~g~~tggaca~~a~~agctactcaagaactgaagacatcaaca
atgctttgccaatgaag~~t~~ctcatggaccgc~~t~~ccgc~~c~~atcttct
actcaagcgacaacaacacagagacc~~a~~gtgaa~~a~~acatatgg~~t~~gc
gatctaatttgc~~a~~agtgc~~c~~tcacaagaggtactgttcaagccat
gg~~t~~atggc~~a~~cgc~~t~~gtgatc~~t~~gc~~g~~atttctggatttgc~~t~~ttgtatg
tttatttctac~~t~~ctaga~~a~~agagg~~t~~caaaaagtt~~a~~tagctt~~c~~ac
cgtgagaatgtt~~g~~ttt~~c~~accagattcatgt~~g~~tatgatgaaaaag
aca~~a~~agc~~a~~acaagagttttcttgc~~t~~tttaggttacaagaacaaga
gtatcg~~t~~tataa~~g~~tcaacaaagattgaa~~a~~acatatttgc~~a~~agg~~g~~
agtgg~~t~~tagaatctt~~c~~ctactcttgc~~c~~ttctactaagacaa
aaaaaagacttgactttgtctaa~~g~~gtttttgtggatattattaacca
agtcc~~t~~tttgc~~a~~aaaagtaatattgttttgcattc~~c~~tcttttag
aatttagtt~~a~~atctagg~~c~~ttatattgg~~t~~tattacttctt~~g~~aaaa
atgatctgtttattctattcatacttgg~~t~~ac~~c~~tcg~~c~~ttttatctt
acttctac~~a~~aaaaggattatc~~g~~taa~~g~~tttagtcttactctcacc
ttccgaaaataaaacaaatatcgata~~t~~cttagatcaa~~a~~cca~~g~~t
tgattaaaacatccctattccctacgattctgatctt~~g~~agatattatt
atcatgtta~~a~~gatctaaattgacaaga~~a~~actgattt~~c~~atttctta
taggaaaaataattactatttagt~~g~~atcatgattgtcgaccgtaaga
gg~~t~~gg~~t~~tagttactctccatcttcttgc~~a~~agaagtc~~g~~aaaa~~g~~tca
gaaattat~~t~~atcaaaat~~a~~acatcaatattgaacacatata~~t~~gtat
gg~~t~~ttatgtttagaaaattccaa~~t~~atttata~~t~~attcc~~t~~tagggaaaa
agaagcttattctcaaattattgttat~~g~~agtc~~g~~ttaaaatatggat
aaaaatataa~~g~~tctaa~~t~~attaaaaactcagtt~~g~~ttgcttgc~~t~~ttta
cctctccaa~~g~~tctccaa~~g~~tcaaatttatt~~g~~at~~t~~taattaa~~acc~~aa
aaaagg~~t~~ttatttagtcaaacttagcatg~~c~~aat~~g~~t~~g~~ttgg~~t~~acccaa~~acc~~
caagcatttagtcttttaatcttctttctccaaataagtttac
aatttttaattgtttgcattccctt~~g~~attattt~~t~~atctcatcccaa
tttagctaataccaaactccg~~t~~tttattcttccaa~~g~~tctttccta
taaatacgttcttcccttatt~~t~~catatcactcaccacaaag
tcttctcatttccctcat .

50
149. An isolated promoter comprising the following nucleotide sequence:

atgttgtgagtgaaggagaagaagagggaaacaaaggatt
tattttagcgagttttgtttgtacgcgggtttgtctgttcaa
tggtgacgaaacgagtgagagagtgctgttattaaagaaaaccct
aattaagtgcagacccgcggttataaaaaatagtcaaaaagttagaaa
acgcgtgtgagtgagacagagacagccattgttgcattatggg
cttataagcgagacgtgttaattgggcttttcattatggccgaaa
acaaaagaaaacgtcgccctgagagattcgaaactctcgccggcagagcc
catgtacttagcaggcacacgccttaaccactcgccaaagcgactt
gttgctatgagtttagacaaaatcattaaaattcttattatgatttc
tcatagtgtgttatattgtggatctactaaaaattcttgcatt
tattacttattttgtgaatttagttgatataggttaagtacaaagtt
aactttattattactcaaaattatcagattaactgattttatatt
gttccctttggtatatagacgtactatagtttttagaaaaaccataa
gattcccttatatttcatagagtgaagagatgagatgagatcttgc
tggagaagaaaataagttccacgaggaggactttttttggta
agacgaggaggaggactttgggtatccagtcttacgttagacat
cgaccctacatttatttgccttcttatcaacatggcaggtaaaa
atcttcattcaaccgaaccaaccaaaatcttccaaataattca
agcaccatccttggaaactcatacataactacagtctacacttt
catttcttcaacgctcaacttaacaaatgatatagtcttagttgc
aattatatgtttatttagtgcatttacatcaaattctgggttgcata
tttgcatt
tatggcgtgatcttataatataacatataatagaatcgttagattat
tttatt
gtttgttatatgataccatattatagttacttttttttttttttttt
gcgataatatatatatcaacttttataacaaaaaaatgataacac
atggtaaagaaaaataaaaaatgaagacatgggtgacacgaaaatgg
cactaaatatacatatataatagatagactacaatatcccatcata
cacttttttaattgactaatacataacttacacacttttttaattga
ctaattcataacttttatttgcattgtcaacatgcaatttatattcc
gttgaactatttatttttttttttttttttttttttttttttttttt
aataaaaaatatgatttccaaatgacgttagagaaaaaaaagaaaa
gttgcgtgtgttttttttttttttttttttttttttttttttttttt
aagtaatataactgcctcctaatttcttgcgcatttcttaccgaagaatc
tctccacttttgccttgcatttttttttttttttttttttttttttt
tttttcaacttttcccaagagaacaatagaaaaaccacacttgc
tcttagggtt
tcatttttggaaagcttaccaccagcgaaaaaaattataacttccatcg
attcctggcttctctctcgctctctgcattgtgcataatcgccg
gactgatcactgtcacctctgtt .

51

150. An isolated promoter comprising the following nucleotide sequence:

52
151. A transgenic plant or transgenic plant tissue comprising an isolated promoter according to any of claims 143 through 150.

54
APPENDIX 1

SEQ ID NO:	INTERNAL IDENTIFIER	FUNCTION OF POLYNUCLEOTIDE / GENE
1, 2, 3	2293133	glyceraldehyde-3-phosphate-dehydrogenase
4, 5, 6, 7	7143495	Histone H4
8 & 9	7143515	ATP dependent RNA helicase, mRNA sequence
10, 11, 12, 13	7143527	nematode specific
14 & 15	7143602	protein serine-threonine phosphatase 1, catalytic subunit
16 & 17	7143612	40S ribosomal protein S4
18	7143666	cytochrome p450
19, 20, 21, 22	7143675	Neuroendocrine protein 7B2
23, 24, 25	7143839	nematode specific
26	7143863	40S ribosomal protein S17
27 & 28	7144016	vacuolar ATP synthase subunit G
29	7144025	malate dehydrogenase
30 & 31	7144060	J2 pcDNAII Globodera rostochiensis cDNA similar to Bystin, mRNA sequence
32 & 33	7144225	similar to arginine kinase
34	7144354	pyrroline-5-carboxylate reductase

SEQ ID NO:	INTERNAL IDENTIFIER	FUNCTION OF POLYNUCLEOTIDE / GENE
35, 36, 37, 38	C10	ribosomal protein L18a
39, 40, 41, 42, 43	C118	ribosomal protein S11
44 & 45	C122	ribosomal protein L16/L10E
46 & 47	C127	FMRFamide-related neuropeptide precursor
48	C129	ADP-ribosylation factor 1
49	C130	ribosomal protein L11
50	C137	nematode specific; conserved in <i>C.elegans</i>
51 & 52	C138	ribosomal protein L7
53	C145	ADP/ATP translocase
54 & 55	C148	troponin
56 & 57	C154	calponin
58	C16	translation elongation factor EF1A
59 & 60	C18	40S ribosomal protein S16
61	C27	ubiquitin
62 & 63	C46	nematode specific
64, 65, 66	C48	ribosomal protein S3AE
67	C59	40S ribosomal protein S5/S7

SEQ ID NO:	APPENDIX 1 (cont.) INTERNAL IDENTIFIER	FUNCTION OF POLYNUCLEOTID E / GENE
68	C8	glyceraldehyde 3-phosphate dehydrogenase
69 & 70	C82	60S ribosomal protein I30/L7E
71	C90	glyceraldehyde 3-phosphate dehydrogenase
72	C135	nematode specific
73& 74	C206	predicted troponin
75	C227	cytochrome P450
76	C238	vacuolar ATP synthase subunit G
77	C246	40S ribosomal protein S4
78	C308	FMRFamide-like neuropeptide precursor
79	C342	ubiquitin
80 & 81	C344	nematode specific; conserved in <i>C.elegans</i>
82, 83, 84, 85	C370	40S ribosomal protein S5/S7
86	C426	nematode specific
87	C458	histone H4
88 & 89	C481	ribosomal protein L30E
90 & 91	C556	nematode specific; conserved in <i>C.elegans</i>

SEQ ID NO:	APPENDIX 1 (cont.) INTERNAL IDENTIFIER	FUNCTION OF POLYNUCLEOTID E / GENE
92	C628	ribosomal protein S17E
93 & 94	C665	malate dehydrogenase
95 & 96	C669	malate dehydrogenase
97	C694	ribosomal protein S3AE
98 & 99	C709	ADP/ATP translocase
100 & 101	C714	ADP-ribosylation factor 1
102	C721	calponin
103 & 104	C726	ribosomal protein L11
105	C736	nematode specific
106 & 107	C773	troponin
108	C834	nematode specific
109	C860	bystin
110 & 111	C863	troponin
112 & 113	C883	translation elongation factor eEF-1A
116	C888	40S ribosomal protein S16
117	C898	glyceraldehyde 3-phosphate dehydrogenase
118 & 119	C935	peptidyl-glycine alpha-amidating monooxygenase
120 & 121	C937	calponin
122 & 123	C942	peptidyl-glycine alpha-amidating monooxygenase

SEQ ID NO:	<u>APPENDIX 1 (cont.)</u>	FUNCTION OF POLYNUCLEOTID E / GENE
124	C954	arginine kinase
125, 126, 127	C969	calponin
128 & 129	7235653	ribosomal protein L18A
130	8005381	neuroendocrine protein
131	7235496	pyrroline-5-carboxyla te reductase
132 & 133	7275710	protein phosphatase pp1-beta catalytic subunit
134	7923685	nematode specific
135	7641370	40S ribosomal protein S11
136 & 137	7923404	nematode specific
138	7797811	ATP-dependent RNA helicase
139	7143613	predicted phospholipase D

Appendix 2:

Exemplary genes used for RNAi vectors.

Promoters:

Constitutive:

Super Ubiquitin from Pine

CCCGGGAAAACCCCT CACAAATACATA AAAAAAATTCTT TATTAAATTATC AAACTCTCCACT ACCTT TCCCACCAACCGTTA CAATCCCTGAATG TTGGAAAAAAACT AACIACATTGAT ATAAAAAAACTA CAITTA CTTCCTAAATCATAT CAAAATITGATA AATATATCCACT CAAAGGAGTCTA GAAGATCCACTT GGACA AATTGCCCCATAGITG GAAAGATGITCA CCAAGTCACAA GATTATCAATG GAAAAATCCATC TACCA AACITTACTTTCAAGA AAATCCAAGGAT TATAGAGTAAAAA AATCTATGTATT ATTAAGTCAAAA AGAAA ACCAAAGTGAACAAA TATTGATGTACA AGTTGAGAGGA TAAGACATTGGA ATCGTCTAACCA GGAGG CGGAGGAATTCCCTA GACAGITAAAAG TGGCCGGAATCC CGGTAAAAAAGA TAAAAAATTTTT TGTAG AGGGAGTGCTTGAAT CATGTTTTTAT GATGAAATAGA TTCAGCACCAC AAAAACATTCAAG GACAC CTAAAATTTGAAGT TTAACAAAAATA ACTTGGATCTAC AAAAATCCGTAT CGGAAATTCTCT AAATA TAACTAGAATTTC TAACCTTCAAG CAACTCCTCCCC TAACCGTAAAAC TTTCTCTACTTC ACCGT TAATTACAITCCITAAGAGTAGATAAA GAAATAAGTAA ATAAAAGTATTCA CAAACCAACAA TTTAT TTCTTTTAAATTACTT AAAAAAACAAAA AGTTTATTATT TTACTTAAATGG CATAATGACATA TCGGA GATCCCTCGAACGAG AATCTTTATCT CCCCCTGGTTTGT ATTAAGGTAATTTATTGGGG TCCAC GGGAGITGGAATCC TACAGACGCGCT TTACATACGTCT CGAGAACGCGTGA CGGATGTGCGAC CGGAT GACCTGTATAACCC ACCGACACAGCC AGCGCACAGTAT ACACGTGTCATT TCTCTAATGGAAATGT CGTTGTTATCCCCGCTGGTACGCAACC ACCGATGGTGAC AGGTCGTCGTGTT GTCGTGTGCGGT AGCGG GAGAAGGGTCTCATC CAACGCTATTAA ATACTCGCCTTC ACCGCGTACTT CTCATCTTTCT CTTGC GTTGTATAATCAGTG CGATAATTCTCAG AGAGCTTTCAT TCAACCCGGG

Strawberry Banding Vein Virus 1

aagctttcactgtgggtaattcattaatctatccagggtggaaaccccaaggaga tctctttctccaaaagaccttacagggcaatcaaaaactacagaaccaggattt gtagtgcacagagtagaccaattacactgagaatcagcgtaccccttagagtggg aaaaatgtgacatccttattccataaccactggattgaggtagactatccaatggaa aaattccatggacaagtcatataagaagaccgcaacagtcgagtatcttccagaga taactgcactcagacctaagataaaagcagtatataatcgtgtactaagatct tcgcagattcaaagaagaagctt

Strawberry Banding Vein Virus 2

Gtttaaacaacagcccaagataacacagaaaagtcaaagggtttcgaaagaccacttgt gactaaggatcatttcatccataattatctggtagcacagactcatgataactgcga ggaacacaagttttacagtgcattcaaaagacactttctttacgggtttcattga aggagccgaccagaatatgtcagagaagctttactgtgggtaatttcatataat ctatccagggtggaaaccccaaggagatctcttctcccaaaagaccttacagggc aatcaaaaactacagaaccagatggtagtgcacagagttagaccaatctacctgag aatcacgagtaccttccatagatggggaaatgtgacatccttattccataaccactg gattgaggtaggactatccatggaaaattccatgggacaagtcatataagaagac cgcaacagtcgagtatcttccagagataactgcactcagacctaaaggataaaagc agtatataatcgtgtactaagatcttcgcagattcaaagaagaagcttaactatgc ttagtgcataagataattctaataagcaattattcagaattatcaaggagaaagaatt aataactctttcagaatatgaagcccgcttacaagtggccagctagctatcactga aaagacagcaagacaatgggtctcgatgcaccagaaccacatcttgcagcagatg tgaagcagccagagtggccacaagacgcactcagaaaaggcatcttctaccgacac agaaaaagacaaccacacagtcatcatccaaacatgttagactgtcggtatgcgtcggtcgaagataagactgaccccaggccagcactaaagaagaataatgcaagtggctcttag ctccacttttagcttaataattatgtttcattattattctctgctttgcctctat ataaagagcttgttatttcattgaaggcagaggcgaacacacacacagaacacctcc tgcttacaaaccatgtatttagctaaacctcttaggaggatata

Nematode Inducible:

Trypsin Inhibitor from *Arabidopsis* (clone#6598343)

ccccggggagcaaaagaacacccagagaagaagaagaaaagcactacagagaaaaatgtg
agcttaagcgctccaaacaacacttctggagtctaaaggatgtcaaaaaaaaa
cttgggtggagacttccgcatatttccaagcatgggttatttttttagcacaca
aactatctgaccctcgacttggattttctgcagttgtccaaactacattgaaac
ggatatgcaggcaacatggatcatgagggtggccatctcgtaagattaacaaagtga
acaggtcactaaggaaaatacagacggtaactggactcggccaagggtttagaaggag
gactaaagttcgactcagcaactggcaattcattgcagtttagaccttttattcaag
aaattgataccaaaagggtctgtcgtctttgataatgatgcacatgcaagaagaa
gtcaggaggatgcctgacgatacttcattcaagctccaggaagctaaatctgtcg
acaatgccattaaagtttagaggaggatacaaccatgaatcaagcaagaccaggtaaga
acttcttatccataaaccatagatggagcatttagaatcttaatccattttcagtt
tttgcaggatcattcatggaggttaatgctagtggcagccatgggcttggatggcc
aaagagtctggctgaatggcagtgaaaggataaaagagcgttgcacacttaagctct
gtggaaatttcagatggaaatggatccaacaatccgatgcagttgcagttgtgaa
cctaacaatccatgtcatgcagcatatcagattcatcaaattggctcaggcgcagtt
ctgcgttggaaagctcatctacttccatggaaagatttggaaaccatgagaacccacaac
agtaatagcagcggagacttcaacaacgcgtatcgtaaaggccaggatagagaa
gacactgtacgttcaagttcgagccatcagttgggtgtctcagcttacaaagaa
gttggaaaacgtttaaactgcaggacgggtcgttcagctgaagtacttggatgat
gaagaagaatgggtgatgttgcgttacagattctgatctccaagaatgatgttggagata
ttacatgttatggaaaacactcggtgaagttctcggtcgatgttgc
ctaggttagttctggcagtaatgttatcttgcacaggcttatgcacgcgt
acatagacacacacagttatgttattcccagtgaaagaatgttgc
tattatgttgcattataataggcatgaaggagaaagacaatggatgttgc
tcagcagaaaatgttatgttttgcgttatataatgc
tgttatatctacgttgcataatgttgcacgcgttgc
agaacacttttagttatccctgtatgc
cctgtggaaaaccaacaaggtaactaaatttgcgttataattgggttgc
aacggaggacttgcatttttagtggcgttggcctataattgttgc
tttcccccttatcagttaaacgcgtccatattccatatttttttttgc
agttcatatccatattttatgc
tttcatatttgcataagggttgc
ctcagtcataatgttgc
ttgttatgggttgcgttatcatca
taaatttatttgc
caaaggacccggg

Arabidopsis Transmembrane Protein from Arabidopsis
(clone#6468048)

ccccggaaattggcactcttcttctgctgggttccaaaagaaaacgaatcaatatgtgc
aacaagaagagctccagaagcagtcatcttctaaaatcttaatctaacaacagctca
agaagaaaaaaattccatagctagagagaacacaaaagtacaagacgacgtcgtaga
ggcacaaggtaaaacctgaatggcttaagccgaactgagtggtttgactagacat
catcagaaaagtctccaagacggtagtcggatgttagatcgctcaagtaattttgg
tttgggttgcacgtttcagctgcccattgattcagttgggtttccctta
tctctaaaggcccaatttcatttaggttagttattgatcattatccctactata
aaggcttcgccttcgagaaatttagggttctgtctgtctgcactcaggtt
tgtgcctcaacgactgctcacttctagcttgcattttcttcgttataatgtat
actgtacattagattattcttgcattcgcagcttcgtatagatttgattcttt
tttgggtgtcttgcattcgcattcgcattcgttgcatttgcatttgcatttgcatt
aaaatgaggtacttgacgcatttgcattcactgtttgcatttgcatttgcatttgcatt
ctctgaatcgtgattcagagacgtattgttcttcgtcatatgcatttgcatttgcatt
agagaacaatacgtctctgaatcgtgattgttttgcatttgcatttgcatttgcatt

**Diaminopimelate Decarboxylase from *Arabidopsis*
(clone#4159709)**

tgctttatgtatggtccggccccggg

Peroxidase from Arabidopsis (clone#4006885)

ccggggcaatcaaggtaacgaaggaggatcagcgaaaggatgggctataattggagt
 ttttcctgcgtgtaaatgtttgtatcttccatgcggacatataactgaaga
 ataaactcaactcattgtttctgggtgtttcttctgtatcagattcctcggtcat
 ctgcactttctgtgtgggggttatttataaaaacaagagtagagcgtgtggtaa
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 cggggatgtcacttctatcgggttactagaaaactgtaaatcacaagaaaactca
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 aacaggtgagtgaaatgttgcacaaagactcaagaactgaagacatcaacaatgctt
 ttgccaatgtaaatgtctatgggaccgcctccgcatttctactcaagcgacaacaa
 cacagagaccaagtgttgcacaaatgttgcgtatctaatttgcatttgcctcaca
 agaggactgtttcaagccatggatggcacgttgcatttgcatttgcatttgcattt
 tgctttgtatgttattttctacccatcataagaaaagaggtaaaaaagttatagcttca
 ccgtgagaatgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
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 tgaaatgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
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 tattgaacacatatactgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 tccttagggaaaaagaagcttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
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 agtctccaaagtctaaatattttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
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 ttctccaataatgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 tcatcccaatgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 aaatacgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 cctcatcccccggg

**Mitochondrial Uncoupler from Arabidopsis
 (clone#4220510)**

ccggggatgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 gagtttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 gagtttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 agtcaaaaatgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 ttatgggcttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 aaagaaaacgtcgccgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 ggcacacgccttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 attaaaatgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 aaattcttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 agttaacttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 ttgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 atagagtgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 aggactcttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 acgttagatgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 aatcttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
 ctgggaaactcataactacactacgtctacactcttgcatttgcatttgcatttgcattt
 acttaacaaatgttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcattt

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tcaaatctgggttcatattgtatgtactatcccgaaacatctcaatgtcccgaa
atacaatcgctatcatatataatcccgatcggttattcttataagatagaataata
tggcgatcttataatataacatatacgatcgtagatttttttttttttttttt
ttatatacgatcataattgcaaaatacttatatatgtttttatataatgataccat
tttatagtttactaaaaaaagttaagcgataatataatataatcaacttttataac
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aatggcactaaatatacatatataatagatagctacaatatccatcatacacactt
ttttattgtactaatacataacttacacacttttttattgtactaattcataacttt
ttatcattgtcaacatgcaattcatattccgttgaactatttttttttttttttt
tttaaaagaagggcttcgttaataaaaaatgtattccaaatgacgttagagcaa
aaaaaaaaaaagggtgtctgtctggtaaaatgaaaaagcaaagcgttggatag
aaaagtataatactgcctcctaatttctcgcccttctaccgaagaatctccact
cttgcctcttcgaaaccctaaaccagaagcaccagatttttcaacttttccca
gagaacaatagaaaaacccaaacttgtctcttagggttttttttttttttttttcatc
tttggatttttttgggtcatatttggaaagcttaccaccagcgaaaaaattataaa
cttccatcgattcctggctctctctctcgctctctgtcatgtctaaatcgccgg
actgtatcactgtcacctctgttcccggg

Stress protein from *Arabidopsis* (clone#6598614)

Pectinacetyl esterase from *Arabidopsis*

(clone#6671954)

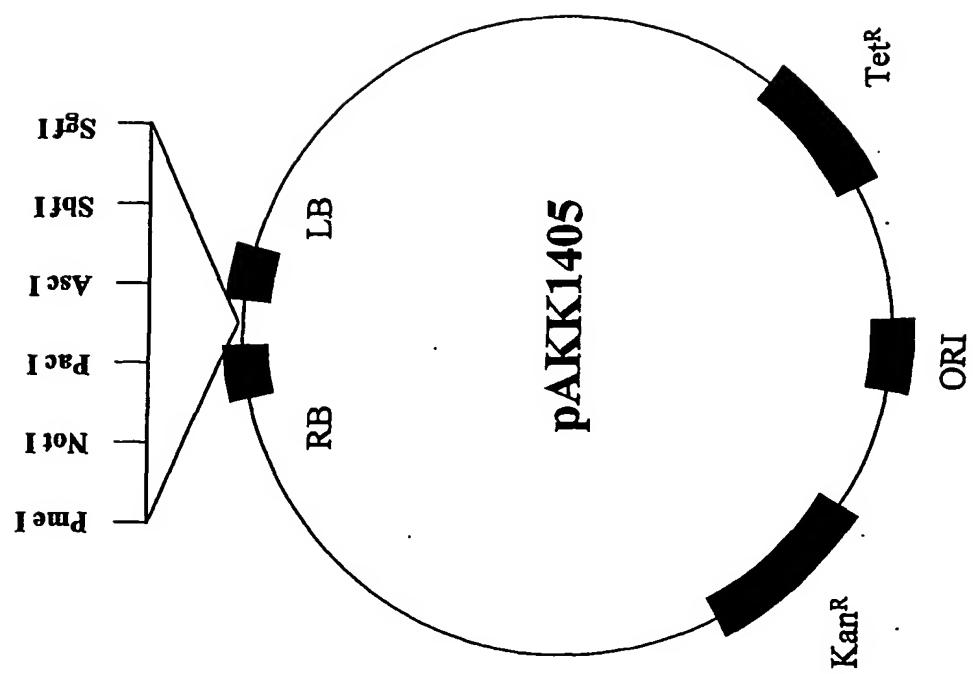


FIG. 1

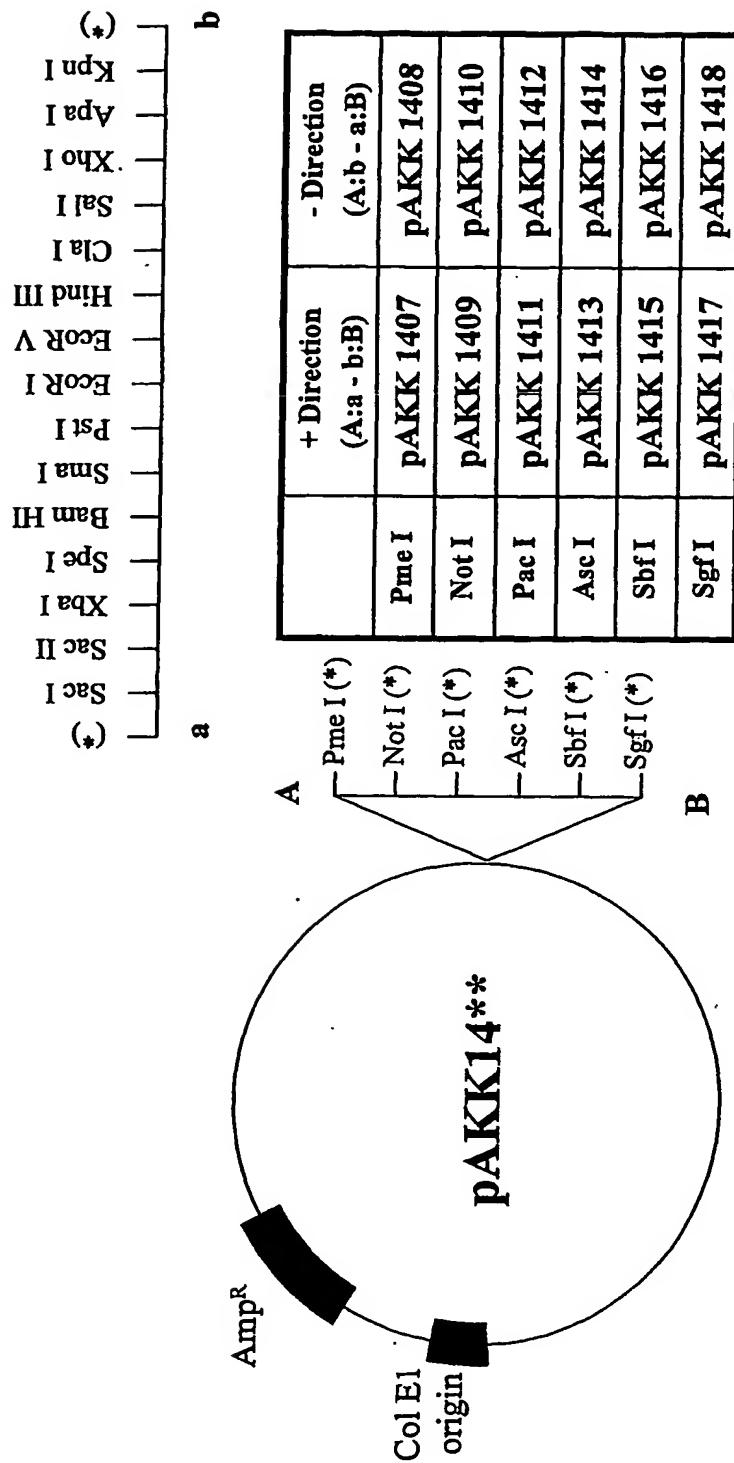


FIG. 2

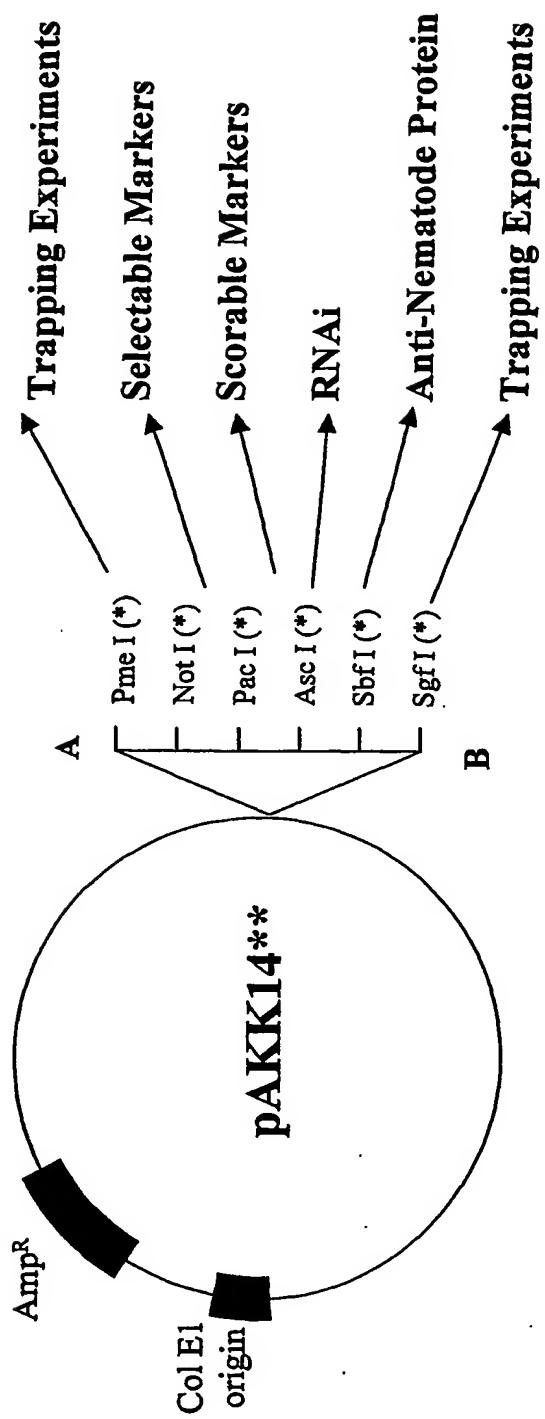


FIG. 3

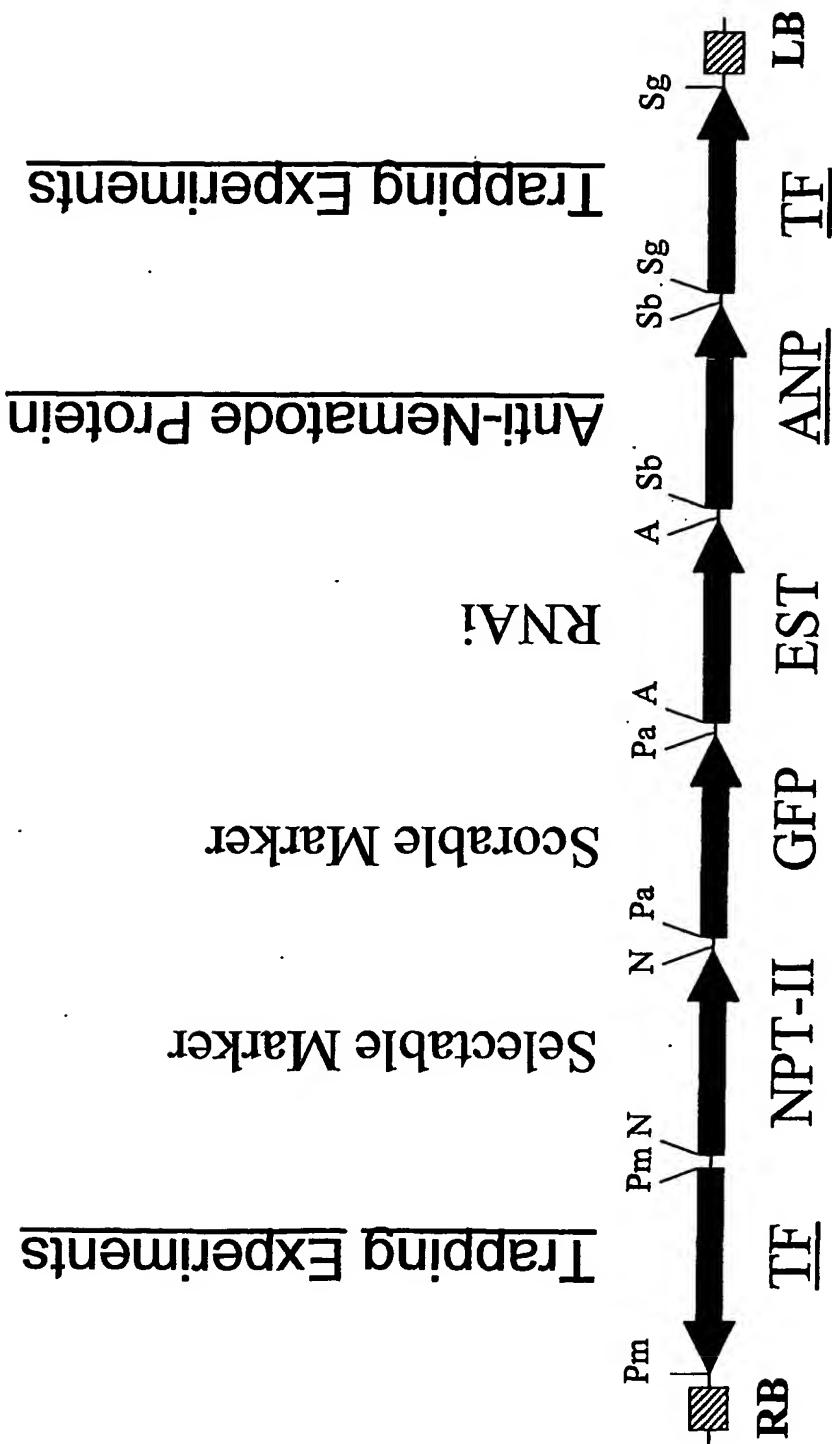


FIG. 4

Selectable Markers

pNOS / NPT-III / tNOS

pSU / Bar / tNOS

pUBQ3 / Intron / Bar / tNOS

pUBQ3 / Intron / PMII / tNOS

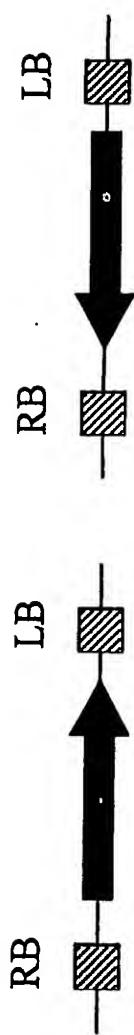
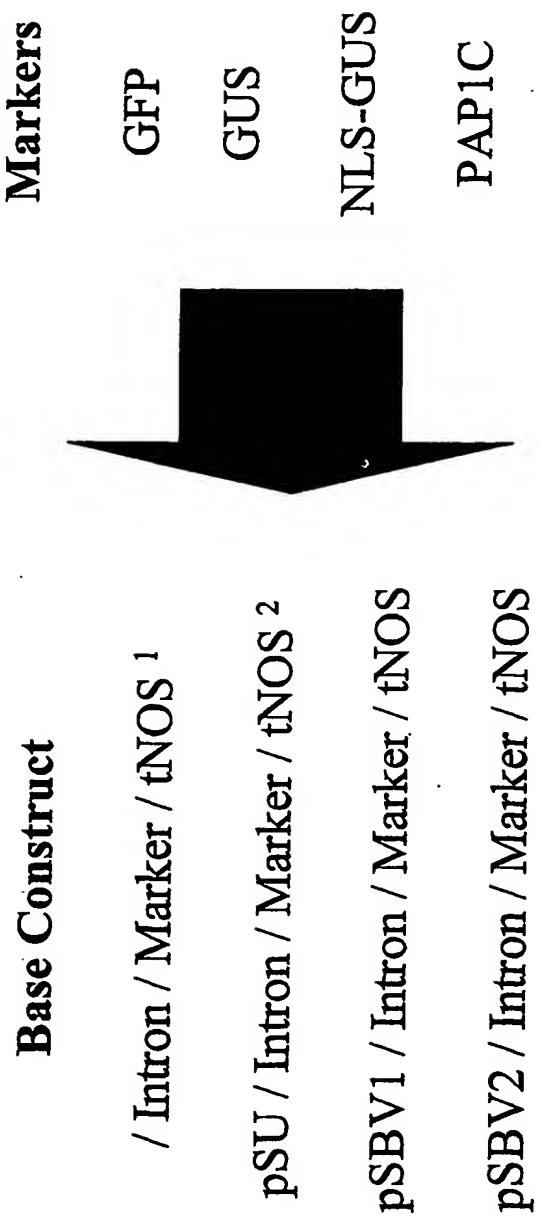


FIG. 5

Scorable Markers



¹ Construct useful for promoter analysis.

² Construct useful for high constitutive expression of genes of interest.

FIG. 6

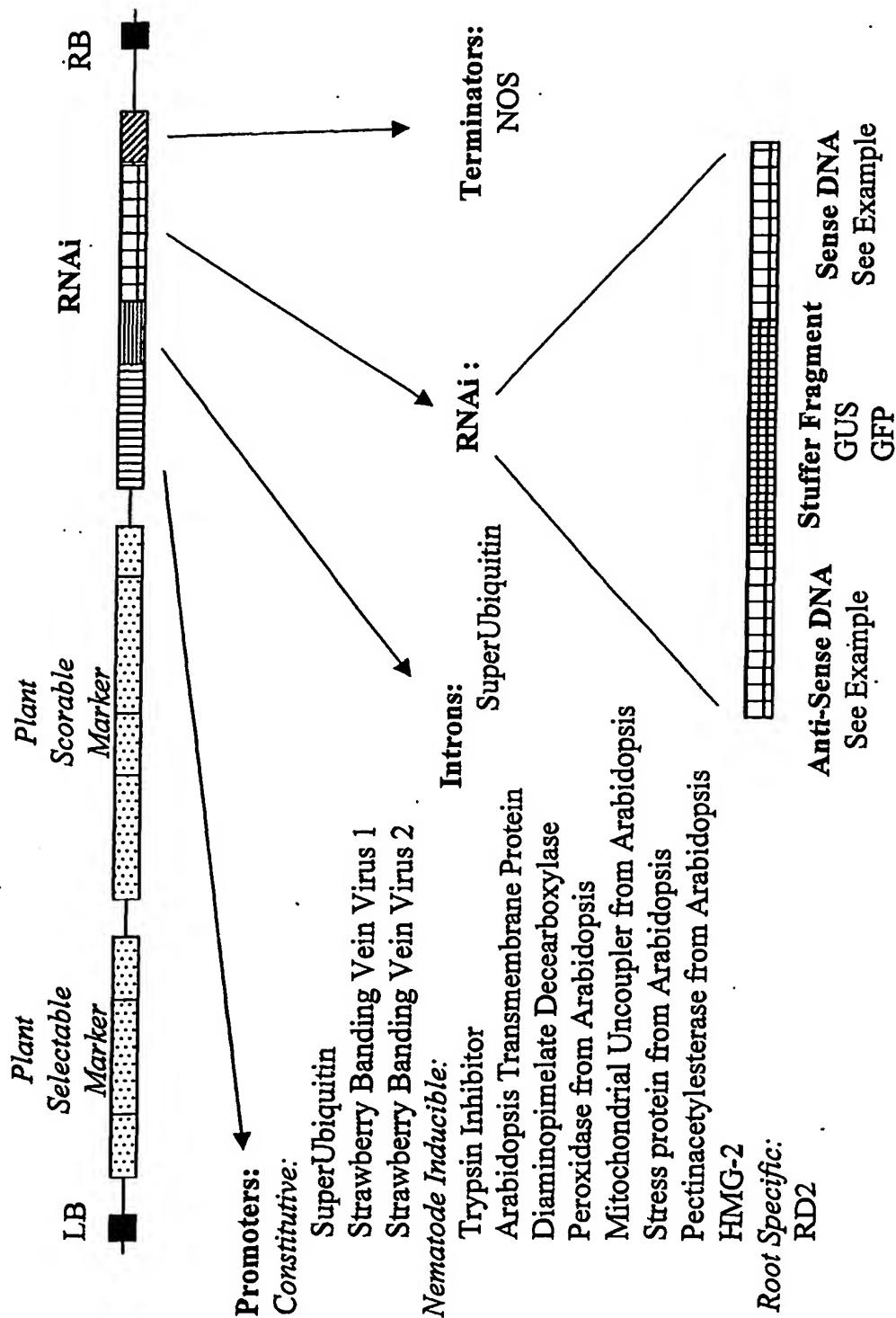
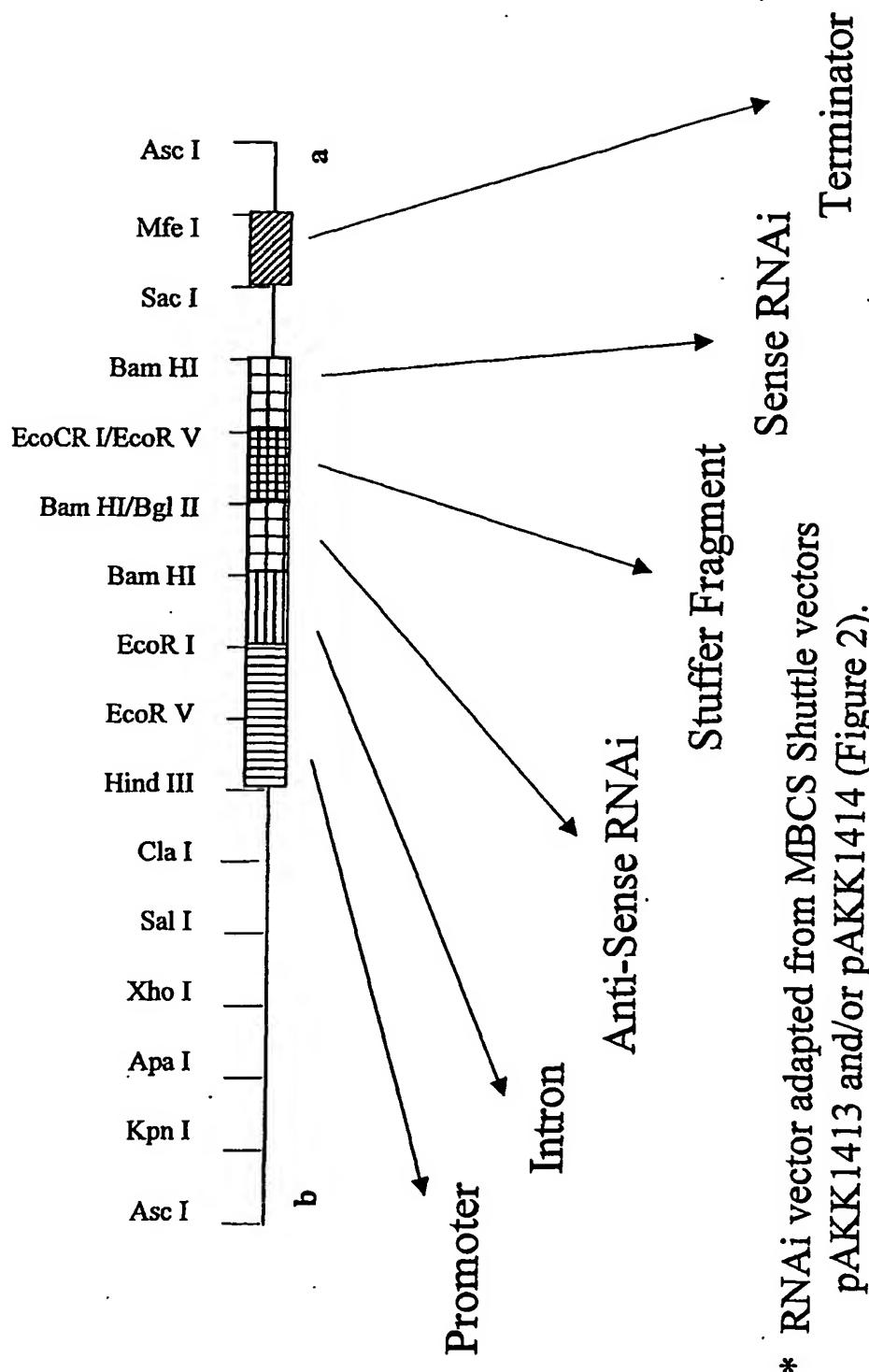


FIG. 7



* RNAi vector adapted from MBCS Shuttle vectors pAKK1413 and/or pAKK1414 (Figure 2).

FIG. 8

AKK110P1
SEQUENCE LISTING

<110> Mushegian, Arcady R.
Taylor, Christopher G.
Feitelson, Gerald S.
Eroshkin, Alexey M.

<120> Materials and Methods for RNAi Control of Nematodes

<130> AKK-110P

<140>
<141>

<160> 139

<170> PatentIn Ver. 2.1

<210> 1
<211> 165
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gcttggcggtt gcgcgctgctg gttgagaagg acaccgttca ggtgg 165

<210> 2
<211> 342
<212> DNA
<213> *Globodera rostochiensis*

<400> 2
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ttcgacaagc gccggcaatt tggcgttga gaaagagggg aaggccacgc acaccatcaa 120
ggtgttcaac ctcaggacc cggccgagat caaatgggcg gaggtggcgc cggaaatatgt 180
gatcgatgc accgggggtgt tcactaccat tgagaaggct tcggcacact tgaagggggg 240
cgccaagaag gtggtcatgt ctgctccgtc cgctgatgca ccgatgtacg tgatggcggt 300
caacgaggac aaatatgacc cggccaagga caacgtgatt ag 342

<210> 3
<211> 205
<212> DNA
<213> *Globodera rostochiensis*

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gaagccggcc tcattggacg ccatcaaggc ggcggtaaag aaggctgccg aagggaaattt 60
gaaggccatt ttgggttaca cagaggacca ggtgggtgtcc acggactttc ttggagacag 120
tcgctcgatcg atcttcgacg ctggggcggtg catctcgatgt aacccgact ttgtcaagtt 180
ggtcagctgg tacgacaatg aattt 205

<210> 4
<211> 167
<212> DNA
<213> *Globodera rostochiensis*

<400> 4
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tcgtccattt gtcattgtg gcccataaga gggccgtttg gtttagttt ttgggtgtcc 120
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<210> 5

AKK110P1

<211> 41
 <212> DNA
 <213> **Globodera rostochiensis**

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41

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 <212> DNA
 <213> **Globodera rostochiensis**

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 cttaacgcct ccacgacgg 79

<210> 7
 <211> 168
 <212> DNA
 <213> **Globodera rostochiensis**

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 <212> DNA
 <213> **Globodera rostochiensis**

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 gccgggacgt ctttggcct taggacgcac tggacatctg aagctgaaag gcgtcaaatc 240
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 gaaatcttc aaaatgacgc ctcaggagaa 330

<210> 9
 <211> 136
 <212> DNA
 <213> **Globodera rostochiensis**

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 tacgtcgacg acgaggctaa gcttacgc 120
 ttcaacttctcc aacaatacta cgttagactg 136
 aaggaaaatg agaaga

<210> 10
 <211> 141
 <212> DNA
 <213> **Globodera rostochiensis**

<400> 10
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 ttgttgtca tcactttttt cagcagcgac aatacggcca atccggtgaa agggccaaag 120
 tcaatagctc gctcggtacc t 141

<210> 11
 <211> 141
 <212> DNA

AKK110P1

<213> Globodera rostochiensis

<400> 11

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 g~~c~~cattttgt ttctacagca cacgcacacc gtcgtctta cagcgttcac ctcgccaaa 120
 aagtagccgt atttgcgaaa t 141

<210> 12

<211> 37

<212> DNA

<213> Globodera rostochiensis

<400> 12

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37

<210> 13

<211> 161

<212> DNA

<213> Globodera rostochiensis

<400> 13

gcgcgttcca tcgcccgcac cacaaaaagt cccatcgctt catatcgtag cgcaaattgt 60
 ctttggtgca aatggcaaaa cggccaaaat aatggtcgaa gccgtacaca accgcccacccg 120
 ccacagcgcc aaccccacac caaatgcgaa atttatcgaa a 161

<210> 14

<211> 306

<212> DNA

<213> Globodera rostochiensis

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 agtttgcata ctcgattgtt agaagttcgg gtttgcgtac cggaaaaac agtgc当地 120
 gacgaatctg agatacgcac ttgtgcatac aaaacacgtg aaattttgt gtcgcagcca 180
 atcttgtgg agtcgaggc acctttaaa atttgtggg acattcacgg acaatataat 240
 gatcttcgtt gattgttcga atatggtggg ttccaccgg aagcgaacta tctatttctt 300
 gggac 306

<210> 15

<211> 261

<212> DNA

<213> Globodera rostochiensis

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 tgaatgcaaa cggagggttcc tcaatcaagt tggaaagac ttcaactgac tgcttcaact 180
 tgctgccaat tgccgcttta atcgacgaaa agatcttttg ctgcccacggg ggctgtctcc 240
 tgatttgcata aacatggcag c 261

<210> 16

<211> 151

<212> DNA

<213> Globodera rostochiensis

<400> 16

gaattctttt agtgcattca gcgtttaatt ttttcgtatt ataataagca tggctcgccg 60
 accaaaaag catttgaagc gacttgcagc accaaaaaaa tggatgtgg acaaattggg 120
 tggcgtttt gcgccacgtc catttgcgg a 151

<210> 17

<211> 306

AKK110P1

<212> DNA

<213> Globodera rostochiensis

<400> 17

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tggacggcaa	agtgcgcacc	gagatgcgct	tcccggtcg	aataatggat	gtgatctcg	120
ttgagaagac	aaacgaaacg	tttgcgtcg	tgtacgtat	gaaggccgt	tttgcgtcc	180
atcgaattca	aaagctggag	ggccagttaca	agctgtgcaa	agtgaagaag	caggccgtcg	240
gggacaagca	ggtcccctac	attgtcacac	atgacgcgc	caccattcgc	taccggaccg	300
ctcatc						306

<210> 18

<211> 528

<212> DNA

<213> Globodera rostochiensis

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ggagcaagca	gacgacctt	cgattggct	ttgttcgtcc	attgggttgg	agcatcgccc	120
gttccctaccg	tatacaaacg	ctgtataaaa	tgaaacaatt	cgattagtca	atttgcgtcc	180
gttcaatctt	agccatttgg	cgcttgaaga	tatgaaattt	ggcaatttta	ttgtgaagcg	240
tgggacacca	attgtaccgc	aggtcagcag	tgttctgttc	gacgaaaaac	tgtatccgga	300
gcccgatcg	tttttgcgg	aacgcgttct	ggacgatgag	ggccgttga	agaaaaagcga	360
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gacattttt	ctgttgccg	ctaatttctt	tctcgctac	aaagtctcc	cgtccgatcc	480
actgaatcct	ccaagcctga	aaaagttggc	ggattatctg	tttacaca		528

<210> 19

<211> 335

<212> DNA

<213> Globodera rostochiensis

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tttggtgatg	gttttgac	ttgcgttcgg	ttgcaccaat	caaattggat	ttgatcagtc	120
ggccgcgtat	ttcccccact	cccagtccat	cgatttgatt	tcgcgcgaca	tcaatccctt	180
ctccggccca	tttggcggtg	gccataaattt	tatgagcggc	ggtgcgggtg	aggcggtcca	240
acagctaggc	cccgaggggc	cctttgagca	gcccacac	gtgaagagt	acaatgttct	300
cccccgat	tgcgagcc	caaattccctg	tccga			335

<210> 20

<211> 52

<212> DNA

<213> Globodera rostochiensis

<400> 20

ggacggctgc acgaaacagt tcgagaacac tgccgagttt tcgcgcagct ac 52

<210> 21

<211> 190

<212> DNA

<213> Globodera rostochiensis

<400> 21

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agcaggatct	ggagcaattt	ctggccaaaca	acggactgca	caaattcaatg	attgccaaga	120
aattccatct	cacgcgggc	gaggagccgc	gccgtcgaaa	acgctttgt	cggccggctt	180
cggccaaaccg						190

<210> 22

<211> 52

<212> DNA

<213> Globodera rostochiensis

AKK110P1

<400> 22
 ccgctacaac ccctacctgg agggcgcccc gctgaagtca gtggccaaaa ag 52

<210> 23
 <211> 54
 <212> DNA
 <213> **Globodera rostochiensis**

<400> 23
 gaattccgac tctcaaggtg gaccacgccc caaccaacag caattgtcag ctgc 54

<210> 24
 <211> 77
 <212> DNA
 <213> **Globodera rostochiensis**

<400> 24
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 aacagaccgg aacagca 77

<210> 25
 <211> 439
 <212> DNA
 <213> **Globodera rostochiensis**

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 acaacgtgca gcagcaacat gttgtggtc aacaacagca gcaacaacag aatttccaac 180
 aaccggccgc cctatcgatc actcacagcc accaacaaca aaaacaacca ccacaagcg 240
 cacagtcgtat gttgtcaatg aaaagtggca atgttgcgt tttgttccg caacaatcg 300
 agcagcacca ctaccaacag cggacactga cgccactgaa gcacacatcc gcacccctcc 360
 cgtccgatcg cttcgatc accaaaacca acagggtgc tccactcccg tcgcagcaag 420
 gcccacggc cactgatg 439

<210> 26
 <211> 539
 <212> DNA
 <213> **Globodera rostochiensis**

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 aacgaaaact gtgaagaagg cgtcgccgt cattatttgc aagtattaca ccaaattggg 120
 cctcgacttt cacaccaaca agcgcatgg cgaggaggcg gccattatcc caagcaaacg 180
 gatgcggAAC cgaatttgcgg gatttattcac acatctgtat aagcgcatgg agctggggcc 240
 tttccatca aatttgcaggaa ggaggagcgc gagcgtcgcg acaattacat 300
 gccccaaatc tcttacctgg atgcgcagaa tcaccagatg atcagcaccc accaagagac 360
 gaaggatgt gcgaaatttc tggggctagg cctcaacttg gaagtggaaag ggcccttgac 420
 gagtggcgcc gctggcgccg gacgtcgatgg agtcaggaca attggcattt ttgtgaaaa 480
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<210> 27
 <211> 179
 <212> DNA
 <213> **Globodera rostochiensis**

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 cggccgaaaa gctgtcgccg gaaaagatta atgatgcccgg gaagcgaaaa gcacagcgac 120
 ttaagcaggc caaacaagaa gcccaggcg agatcgagca gtatcgncag gagagggag 179

AKK110P1

<210> 28

<211> 133

<212> DNA

<213> **Globodera rostochiensis**

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gtcgctggag	gcaatgaatc	gcaatgtcgc	ggcgaacaaa	cagcaggtca	ttgtacgtct	120
gctgcagttg	gtg					133

<210> 29

<211> 482

<212> DNA

<213> **Globodera rostochiensis**

<400> 29

gaattcgtga	aatcaaaggc	tttttaattt	tatttacaca	aaaaatggtt	ccaccaccaa	60
ttcgctgtt	gtcacttgtt	gccgctggac	aaattggcta	ttcactgggtt	ctgcaaatcg	120
caaaaggcga	tgtgttggc	aaagatcagc	caattgttct	cgttctccctc	gacattccac	180
cgatggccga	agtactcttc	ggtgtccattt	ttgaattgtat	ggactgtgcg	ttggcaaacc	240
ttggccgtgt	ggaggcgtgt	accacggaag	agcaggccctt	caaggacattt	gactacgcctt	300
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atgtcaaaat	tttcaagtcc	caaggcgaag	cattggcccg	cttttccaag	cccgtnctgc	420
aaagttctcg	tggtgggcaa	cccgccaaac	acgaacgcgt	acatttgcgc	aaaatatgcc	480
99						482

<210> 30

<211> 605

<212> DNA

<213> **Globodera rostochiensis**

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gccatttctt	ttatcctacg	tgttcttgc	gaaaaaaaaattt	acacacttcc	tttccgagca	480
tttagacggcc	tcgttttca	ttttcttgc	atgcgtcac	atcaggcga	gctgcccagt	540
atttggcacc	agacactgtt	ggctttgtc	gagcgttacg	caaaagacat	aagtgcagaa	600
cagag						605

<210> 31

<211> 112

<212> DNA

<213> **Globodera rostochiensis**

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ccatccccat	catcaaattt	ccccgatttt	ctgcggcttt	tgcgcggcgc	cgagtcgagg	60
aatgaggaaa	gtgaagcaaa	tgtgcccgtt	tatgcgcgt	atgatgaaat	gg	112

<210> 32

<211> 105

<212> DNA

<213> **Globodera rostochiensis**

<400> 32

gaattcgttt	gagcatttat	ttgacaaaat	ctgaataaaat	ggccgtacca	aaagaagtta	60
ttgacaaaat	cgaggcgggt	tacaagaagc	ttcaggaagc	gtctn		105

<210> 33

AKK110P1

<211> 425

<212> DNA

<213> Globodera rostochiensis

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aagaagtacc tcaccaagga agtcgtcgat gcctgcaagg ataagcgcac caagcttgg 60
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 atccctt 425

<210> 34

<211> 581

<212> DNA

<213> Globodera rostochiensis

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 gagggaaatc ttgcttattt cattgattag gaattacact t 581

<210> 35

<211> 102

<212> DNA

<213> Globodera rostochiensis

<400> 35

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 cccatcaaag catccggaga aacattaagg aagtttattt tc 102

<210> 36

<211> 34

<212> DNA

<213> Globodera rostochiensis

<400> 36

tgcaaatgt gcaaacccca cgcttcacaa gatg

34

<210> 37

<211> 100

<212> DNA

<213> Globodera rostochiensis

<400> 37

tcatgttgcg gccaaatctc gcttctggta ctttacgagc atgctgcgtc gagttaagaa 60
 aacacacggaa gagatcggtt cgtgtcaaga ggttttcgag 100

<210> 38

<211> 176

<212> DNA

<213> Globodera rostochiensis

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 gcgagtatcg ctgatgttac cgaggccggt gccgtgaccc aatgtatcg cgacatgggc 120
 gctcgtaacc gcgctcaggc ggatcgaatt caaatcatca aagtcaaacc ctcaag 176

<210> 39
 <211> 155
 <212> DNA
 <213> **Globodera rostochiensis**

<400> 39
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 agcgcgcgtt caaaaacaa ccgatgttt ttctgaacga caagttcaga acgcaaggga 120
 ttggaaagaa ggcattccaa aaggaccgtt actgg 155

<210> 40
 <211> 35
 <212> DNA
 <213> **Globodera rostochiensis**

<400> 40
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<210> 41
 <211> 70
 <212> DNA
 <213> **Globodera rostochiensis**

<400> 41
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 gcggacgatt 70

<210> 42
 <211> 85
 <212> DNA
 <213> **Globodera rostochiensis**

<400> 42
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 cgtgttccg agatgtctct ctcgg 85

<210> 43
 <211> 193
 <212> DNA
 <213> **Globodera rostochiensis**

<400> 43
 agttcggttc aatgtgctca aggtgatcaa agcatcgggc tcgaagaaaag cgttcgacaa 60
 attctgatgc ggccaagcca acccgcaacg gtcatttgtt atggttccta attgttgctg 120
 ttttcaatt atttgtgtta aatgactgaa ttatgatca acggataact agtattcttc 180
 taaaaagct cga 193

<210> 44
 <211> 219
 <212> DNA
 <213> **Globodera rostochiensis**

<400> 44
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 gaagacgtcc ggcgcgttgtt atcgctata ttaagaacaa gccgtatccg aagtcgcgtc 120
 tttgtcgccg tgtacccgac caaaaatttgcatgc gcatggat tttgggtaga aagcgcgc 180
 ccgttgcacga attcccatgc tgcgtgcata tgatatcga 219

AKK110P1

<210> 45

<211> 489

<212> DNA

<213> *Globodera rostochiensis*

<400> 45

tccgaggcgc	ttgaggctgc	gcgaatttgt	gcgaacaaat	atatggtaa	gaattgcgga	60
aaggacgggt	ttcatatgcg	cgtcagaatc	catccatacc	atgttaattcg	catcaacaaa	120
atgttgtctt	gcgctgggtc	ggaccgtctg	cagactggga	tgcgtgtgc	gttcggaaag	180
cctcaggagc	tcgtggcgcg	tgtcgcac	gtgtatatgc	tgtatgcgt	gcgtattcg	240
gaccaacacc	aagctcacgc	attggaggcg	ttccgtcggg	ctaaattcaa	gttccctgg	300
cgtcaataca	tcgtcttgc	ccgcaagtgg	ggcttcacca	aattcgatcg	cgaggatatac	360
gagaaaatacc	gcaaggaggg	ccgtgttatac	cctgacgggt	tgcattgca	gttactcaag	420
caacacggac	ccgctgaagg	agtggctcaa	gaacccatt	taatctctg	tttgtcttg	480
gactcttgg						489

<210> 46

<211> 101

<212> DNA

<213> *Globodera rostochiensis*

<400> 46

gaattccccg	gctcgagccg	ggttgacgat	gtcctccccc	acccctctc	actgcgttcc	60
gtcctccccc	agccggaaat	tgttcctgtg	gctgttgcgg	g		101

<210> 47

<211> 485

<212> DNA

<213> *Globodera rostochiensis*

<400> 47

tccacccaaag	tccattcgct	gtcgcgcagtc	catttattcc	acaaaaaagat	gattccgtcg	60
tcgttccgat	gacgtcggtt	ggccaaaccgt	tgcccccgtc	accgcgttca	ctgggtgcca	120
acccgcgcgt	ttatttgtg	ttcccgagaaa	acttgccgtt	ggagcggccc	ttcgacgagc	180
aaaacgcacgg	ctcccgaggag	gaatttagccg	aagaagcgt	ggaaacgaaag	gcaagagaggg	240
cgcaaacgtt	cgtccgatcc	ggcaaaaggcg	cgcaaaacatt	tgtgcgggtt	ggaaagcgt	300
cacaacatt	tgtacgcctc	ggaagggaca	cgcaaaaggca	attcgatggg	aaaatgcaaa	360
gtgaacagca	acagaaaaaa	gcttaaagca	aacggcgccg	acttttctt	taatgaatgc	420
gcccacccg	catgacaatt	cttttgtgt	atgtgttgc	attttatga	tcggtaatgt	480
taaca						485

<210> 48

<211> 651

<212> DNA

<213> *Globodera rostochiensis*

<400> 48

atctgttcaa	gggactgttc	ggcaagaagg	aatgcgcatt	tctgtatggtt	gggttggacg	60
ctgtggaa	gacgaccatt	ctgtacaagt	taaagctcg	cgaaattgtc	accaccatcc	120
caacaattgg	cttcaacgtg	gaaaccgtcg	aatacagaaa	catctcggtt	actgtttggg	180
acgtgggtgg	tcaagacaaa	attcgccac	tttggaggca	ctacttccag	aacacgcaag	240
gactgtatcc	cgtcggttgc	agcaacgtat	gcccgcgtgt	gggcgaggcg	cgtgaagagt	300
tgtatgcgtat	gctggcgag	gacgagttgc	gcccgcgtgt	gttgctgtt	ttcgctaaaca	360
aacaggattt	gccgaatgcg	atgaacgcgc	ccgaaactgac	agacagactt	ggactgcaca	420
acttgcggaa	ccgcaattgg	tacatccagg	ccacctgcgc	gacttcgggc	gacggactct	480
acgaggact	ggactggctg	agcaaccaggc	tcaagaacacg	aggctaagct	gggttgggt	540
ctgttgcact	tgcccccgccg	attgtatgc	attgaattta	tttgtgtt	tcgcgcgc	600
gctttttgt	gggacgtcg	attaattttt	tattccgtt	t		651

<210> 49

<211> 660

<212> DNA

<213> *Globodera rostochiensis*

AKK110P1

<400> 49

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gaattcccaa gtttgagatc aattcagttt cacttagaca aaaatgccgc cgaaattcga 60
cccaactgag atcaaaaatcg tgtacctgcg ttgcgtcggt ggtgaaattg gtgcaacatc 120
tgcacttgc caaaaagggtg gcccacttgg attgtcgccc aaaaaaattg gtgaagacat 180
tgcgaaggcc acacaggact gggaaagggtc taagggttacc tgcaagctga caattcagaa 240
tcgtgtcgcc aagatcgacg ttgtccccatc ggccgcctct ctgatcatca aagagttgcg 300
cgaacctccg cgagaccgca aaaaagtcaa aaacgtgaag cacaatggca acctgaccat 360
cgagcaagtg atcaacattg cgcgtcagat ggcgcctcg tcaatcgac ggaagttgca 420
gggcaccgtg aaggaaattt tgggaaaccgc ccagtcggtt ggctgcacca tcgatggaca 480
acatccgcac gacattgtgg acgcgatcag agggggagac atcgaatac ccgaggaata 540
aagaaaggac ggcgcctccg atttttgtgg gacggacatt gggaaatttga ggtgaatgag 600
ttgccaattt cattcattca tcaattgttg ttattgntgg tacggataaa tttgttaattt 660

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<210> 50

<211> 625

<212> DNA

<213> Globodera rostochiensis

<400> 50

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gtggcaatcc gggacggcgt cccctacccc ccactgcctc ctacaaaccg atcccccgaa 120
tacatgaaca tgctgaccccg ctccctctcc gtgccaattt tccgcata tctggcgccc 180
atcgaccgt acagacccgtt gttgcccgtg tacacttaca acacttacca cgggtacttc 240
ccctaccgcgca actaccgcgg ctacacccctt gcaatgcgtt actggatcga ccgatactat 300
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aaatggttcg actacgacaa ccctcccaat taccggccct actacaacca tcgccttaac 480
ggatatgctc ggccgtatca ctaccggtcc catgcgttgg cccaccgtt caattacccg 540
gaagaatgg tcagggaaacg ggtctgacaa atcgaactgc tccaaatttga cgtggccgc 600
attcgaagaaga agacgaaaaa agctt 625

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<210> 51

<211> 402

<212> DNA

<213> Globodera rostochiensis

<400> 51

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cgaaactttt gctcaagcga cgcaaaatca gagctgcgcg aaaggccgca aaagcaaaga 120
acaatttgag ttctatcaaa aaagcacggc ccaagaagggt gggaaatcttc aaaagagccg 180
agcagtattt ggtggagttac cgtcagaagc aacgcattt gcttgcgtg aaacgtgaat 240
cgaagaaagt cggcaattat tatgtgcccag aagagccca aactgcctt gtggtccgaa 300
tcaaaggcat caataagatt catccgcgtc ctgcgaaggt tctgcagtt ctccgccttgc 360
gtcagatcaa caacggcggtt ttctgtaaagt tgaacaaggc ga 402

```

<210> 52

<211> 433

<212> DNA

<213> Globodera rostochiensis

<400> 52

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ccgaccggta catcgcttgg gtttatccga gtcagaagat catccgtcag ttggtctaca 60
aacgcggtaa cgccaaagag aaggacagc gcattccaaat aacggataac aacattgtt 120
agcgcagttt gggcaagcat gacgtgattt gtgtggagga tatgatccat cagatttgg 180
ccggtcgac ccgcacttcaa acaggtgacc aacttcctat ggccttcaa gctgagcaac 240
ccgggtggcg ggttcaagaa gaagttcaat cactttgtt gaggggaggcg attatggaaa 300
ccgcgaggac caaatcaaca aattatttgg aagaatggtc taatggaaagg gaagcggana 360
aagaaaggaa attgnggctt tttctgttg ttgttttgac gataaaatttga taactccaaa 420
aaaaaaaaaaa aaa 433

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<210> 53

<211> 768

<212> DNA

AKK110P1

<213> Globodera rostochiensis

<400> 53

gaattcgttt	gaggtaaac	tttattagcg	tatthaacaa	tgtccgaagg	aggagcggaa	60
aagagtagca	gcgggtccaa	gggggggttt	gatgtcaaga	aatttgcgt	cgatcttgcg	120
tccgggtgt	ctgcggccgc	tgtctccaaa	actgttgtt	ctcccatgt	acgtgtcaaa	180
ctcttgttgc	aggtgcaaga	tgcttccgt	cacatcactg	ccgacaaacg	ctacaaaggc	240
attattgacg	tgcttgtccg	tgtgccgaaa	gagcagggt	ttctgtca	gtggcgtggg	300
aacttggcca	acgttatccg	ttatttccc	actcaagcgc	tgaacttcgc	tttcaaagac	360
acccacaaac	gcatctttac	ggagggactg	gacaaaaaca	agcaggctg	gtcgttctt	420
gtcatgaatt	tggcctctgg	aggtgccgg	ggcgccacgt	cgctgac	tgttatccg	480
ctgggacttt	gcccgtacgc	gttggcccg	tcgatgtccg	aaaagctgt	tcccgcgagt	540
tcaacggttt	ggcccactgc	atcgaaaaaa	tcttcaagtc	ggacggtccc	atcggtctt	600
accgggctt	cttcgtctcc	gtccagggca	tcatcattt	ccgcggccgc	tactttggat	660
gctttgacac	cgcgaagatg	atttcgcgc	cgatggcaa	gcagatgaat	ttcttcctca	720
catgggccat	cgctcagg	gtcaccgtgt	cgtccgggt	cctctcct		768

<210> 54

<211> 338

<212> DNA

<213> Globodera rostochiensis

<400> 54

gaattccagc	agattaattt	gaatggctga	gaacatcgaa	gagattctt	ccgaaatcg	60
cggctcccaa	attgaggagt	atcaacgctt	tttcgacatg	ttcgaccgcg	gaaagaatgg	120
ttacattatg	gccacccaaa	tttggacaaat	tatgaacgcg	atggagcagg	actttgacga	180
aaagaccctc	cgaaaatttga	tccgcgaagtt	cgacgcggac	ggttcggca	aactggagtt	240
cgacgagttc	tgcgcgttgg	tgtacacggt	ggccaacact	gtggacaagg	acactctgcg	300
aaaggagctg	aaggaggcat	tccgactctt	tgacaagg			338

<210> 55

<211> 267

<212> DNA

<213> Globodera rostochiensis

<400> 55

gaaattgcgc	ccgatctcag	cgacaaaggat	ttggaggcgg	cggtcgacga	aattgacgag	60
gacggcagcg	ggaagatcg	attcgaggag	ttctgggagt	tgtggcggg	cggaaaccgac	120
tgagaaaaaga	gcaaattcgat	ccaaatccaa	acggacccgt	ccatttcac	ctccatccgt	180
ccgtcgatt	attatattt	ccagtgaaat	tttcccatta	aaattcggt	aaagtaaaat	240
aatttgcga	aaaaaaaaaa	aaaaaaaaaa				267

<210> 56

<211> 597

<212> DNA

<213> Globodera rostochiensis

<400> 56

gaattcgctg	gacacttcgc	atccggagta	cagccacgag	cagagcatcg	accagaccag	60
catccccctac	cagatgggtt	cgaacaagta	cgcctcgca	aagggcatga	ccggctttgg	120
acaccccccgt	tgggagggtc	ttgaccgc	cattctcgat	cagaaccgc	agtgcgaagg	180
aatggttcg	ctacagtcgg	gtaccaaccc	gttcgcctcc	caggcgggca	tgaccggctt	240
cggcacaccc	aggaacacca	cctatgaggc	ggaggcaggc	gagctccct	acgaggacat	300
gaagaagtcg	gaggcgtatc	tcccgtccca	ggccgggttgg	aacaaggcg	actcgacaaa	360
gttgatgacc	aacttcggca	cgccccgtaa	caccaccacc	aaggtcaaag	tggagaattt	420
ggcgaaattt	ccggaggaca	ttttgtctaa	aggacacggc	gaggtgcgc	tgcagtccgg	480
taccaacccg	ttcgcgtccc	agaagggtt	cgtcgcgttc	ggtaccggac	gtgacgtgt	540
ccgtgagggg	gtgaacgtga	acgtgcgtcc	gggcgacttg	gagccgcttc	cgagga	597

<210> 57

<211> 80

<212> DNA

<213> Globodera rostochiensis

AKK110P1

<400> 57

ggcattgtgc gtctgcaagc cggtaacgaac aagttcgact cgcagaaggg catgaccctt 60
ttcggtaggg gccccgtcg 80

<210> 58

<211> 513

<212> DNA

<213> Globodera rostochiensis

<400> 58

gaattcgcca caccgctcac atcgcgtgca aattcgcgcg acttaaagag aagggtggacc 60
gnccgtctgg caagaaagtt gaggacaacc cgaagtcgtt gaagactggc gacgcggaa 120
ttgtcgaact gattccgacc aagccgatgt gtgtggaggc attcactgtac tacgcaccgc 180
tcggccgttt tgctgttcgc gacatgaggc anactgttgc cgtggcgcg atcaaatcag 240
tggagaagac ggaaggcggtt ggcaaaagtga ccaagccagc gcagaagggtc ggcgcgactg 300
gtggcgggaa gaagacatga ccaaggggag gggcggttcc ctaagggcca accgtcgacg 360
aaaatgcac caaccttgc tttatcgat tcttatttgc ttccttccac ccgtctctat 420
ccatattgtc gttgcgttgg ataatgtttt atttttgtt attgtcctgg ttggaaaata 480
aatttggta attaaaaaaaaa aactcgtgcc gaa 513

<210> 59

<211> 393

<212> DNA

<213> Globodera rostochiensis

<400> 59

gaattcgttt gagcgaaaaa aacatactat acaatggcaa caactgagaa gcctcagggt 60
gttcaacagc ccgtgcaggcttggccga aagaagacag caacagccgt tgcgtttgca 120
aaaaggggca agggcttgc ttaaggcattt gggcggttcc tggactacat gcagccggag 180
attcgtcgca ttaagctcca ggagccattt ctcattgttgc ggaaggacaa atttggggaa 240
atcgacatac gaatccgcgtt caagggcggtt ggacacattt cgcaatttgc tgcaatttgc 300
caagcaactgg ccaaggcact ggtcgcttgc taccagaaga atgtcgacga gcagagcaaa 360
aaggaaactga aggacattt tggcgttac gac 393

<210> 60

<211> 154

<212> DNA

<213> Globodera rostochiensis

<400> 60

cacgagccaa agaaattcggttggacccggg agctcgcgct cgctaccaga atcgtaccgt 60
taagaaataa ttttgcgtat caaatgtttt gatgtatgtt cttgtttttt ttgttataaa 120
aaaaaaattttttaaaaaaaaaa ccgcccatac tgac 154

<210> 61

<211> 666

<212> DNA

<213> Globodera rostochiensis

<400> 61

gtattccaag tttgagcgat cagagttctt caatcttattt tcaactgtttt tccatcaacc 60
aactgtcatc atgcaattt tcgtcaagac gtcaccggc aagaccatca ctctcgagggt 120
cgaggcttagc gataccatcg agaacgtgaa agccaagatc caggacaagg agggcattcc 180
gcctgatcag cagcgctgtt tcttcggccgg aaaacagctt gaagacggac gcaccttggc 240
cgactacaac atccagaagg agtccactt ccattctcgat ctgcgtctcc gtggcggaaat 300
gcaatattttt gtcaagacgc tcaccggcaa gaccatcact ttggaggtcg aggcaggcga 360
caccatcgag aacgtgaagg ccaagatcca ggacaaaggag ggcattccgc ctgatcagca 420
gcgtctgatc ttcggccggaa aacagctcgat agacgggcgc actctggccg actacaacat 480
ccagaaggag tccactctcc atctcgat tgcgtttcgat ggaggagaga actgaatcgc 540
gggctgatgg aaagatgacg aatatgtatgtt ctattcgatg acttgcgtctt ttcgtatataa 600
ttgattgtgt tccatttgc ggtcatcaaa tctttatgac cccctcattt ggcattggaaac 660
gataaa 666

AKK110P1

<210> 62

<211> 213

<212> DNA

<213> **Globodera rostochiensis**

<400> 62

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gaattcgttt gaaaaacttt ttcaaccatt cattcaaatg tctcatcaag tgacacgggc 60
agcaactcaac cacgggacgc gtgtactgag cgtgttggag aaggtaagt tggtctgctg 120
gttgaggag acacattcgt tcgcgaagt ggctcgaaga taccgggcag aatttggtat 180
ggaaccacccg cagttggacc aagtgaagaa gtt 213
```

<210> 63

<211> 488

<212> DNA

<213> **Globodera rostochiensis**

<400> 63

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ggcttggag aacaacagcc aattcccgtc gtaagcgatg cgggacttggaa tgcggaagaa 120
cagctgagaa tggccagaat gtgagccgga ggacctgaag atttatgaac gaaattttcc 180
agtgaagtgg accaacgcctc ttcgacttta tctgctttgt gtaaagtgtt tagaatcgcc 240
ttccaattca aaggcttttcc attccccaac ttttattttt ggcgcaaaaaaa ttctttagga 300
taagcgtgaa taatttatttgc atttgcattttt tctttctttt atctccgcct cgaagtcgca 360
agtgttcctt ttggcccggtt ccctttgtt ttgaatgtt ttccattccc atcccctcac 420
tttctcatat ttgtgacatt cagctgcatt gttcgactcc cattttaaag ttgagtgaaa 480
tgcatttgc 488
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<210> 64

<211> 249

<212> DNA

<213> **Globodera rostochiensis**

<400> 64

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wccrgakbng aacahcdkdg vhwatnvcbn gschvbwagc rntcsvddb wgnhnsswtg 60
gkgdyrbwnt msnwrmanrg artsstsgaa ttcccaagggt tgagagtaaa tattattagc 120
taaaaaatggc agtcggaaag aataagagaa tgggcaaaaaa gggagccaag aagaaggctg 180
tcgatccgtt cacacgcaaa gaatggtacg acatcaaagc gccggcgatg ttcacacatc 240
gaatssts 249
```

<210> 65

<211> 362

<212> DNA

<213> **Globodera rostochiensis**

<400> 65

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wcbcrbhdyb ytsgcrsnck tbdsbhcsy gcdwkmtnvk hscngdckty nyykkvbmr 60
ntmsnwrman rgartsstsg tcaaccgtac tcagggaaacg cgcatccgaa gcgactttct 120
aaaaggccgc gtttacgaag tgtcaactggg tgaccttaac agcaactgacg ccgactttcg 180
aaagttccgc ctgatctgtg aagaggtaca gggcaagatt tgcctgacca actttcacgg 240
aatgtcggtt actcgggaca aactgtgctc tattgtcaag aagtggcaca cgctcattga 300
ggcgaatgtg gcagtgaaga ctaccgacgg tttcatgctc cgacttttt gtatcggtss 360
ts 362
```

<210> 66

<211> 128

<212> DNA

<213> **Globodera rostochiensis**

<400> 66

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aatcaaatta agaagacgag ctatgaaaaa gcctctcagg tgcggatgat tcgtgccaaa 60
atggtggaga tcatgcagaa agaggtctct tccggcgatc ttgaangaaa gtagtcaaca 120
agcctgat 128
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AKK110P1

<210> 67

<211> 502

<212> DNA

<213> *Globodera rostochiensis*

<400> 67

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gaattccatt aaaaaactaa acgaacaaat ctaaagatgg ccaccgaagt ggaggaaaat 60
gttcctacgg ttgacccatg ggggtctgtg gaggaagtgg gtggtaaga gtcgatgcag 120
ttggtcagcc ttgacgttac cgaggtcaaa ctgttcggaa aatgtccct taacgtatgt 180
gaagtgtccg acatttcgct tgtggattat attgcggta aggaaaaggc gccaaatata 240
ctggccgaca ggcgcggccg ttaccaacag aagcgcttc gcaaggccac ctgtccggtg 300
gtggaaacgt tgcgtttgtc aatgatgtat cacgggcgaa acaacggaaa gaaactaatg 360
gcgggtgcga ttgtgaaaca ccccttcgag atcatcacct gctaccggag agaaccagg 420
ccaagtgttgc tcaatgttg tgataaacag tggggccnc gaagattnca cacgtatcgg 480
acgtgcgggc actgttcgtc ga 502

```

<210> 68

<211> 519

<212> DNA

<213> *Meloidogyne incognita*

<400> 68

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gcaaaacttt atcaaataaa aaatttatat ttgccaaaca aatttatgaa taaaattca 60
ttaatcatta aaactacatt taaaatatac ttttttagaga atgtcgctca aaatattctt 120
ttctccctt tatgcatcta tctaaccaga cttggaaagca atatggctaa tcaagtcaac 180
aatacggcag gaatacccaa actcggttac ataccagctt accaatttaa caaaatgcgg 240
gttggaaacc ataagaggct cggcgtcgaa aatagacgaa tgagtgcgc caagaaagtc 300
ggtagaaaca acctggtcct cagtatatcc aagaatccct ttaagtttc cttccgaagc 360
agtcttaatt gcattcttaa tagcttcctt cgttgctggc ttctccaaac gagcagtcaa 420
atcaacaacg aaaacgtttg ggcgtcggca cacgaaaagc cattccggt aagcttccca 480
tccaattcat ggattgaccc ttccaacagc ctttgacgc 519

```

<210> 69

<211> 218

<212> DNA

<213> *Meloidogyne incognita*

<400> 69

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ttgattctt attagtggac aatgacggaa gaccagaaga agttgccat ggtgccttag 60
actgttttga agcgaaggaa agttagggct gctcagcgtg cttctctact caagaataaa 120
ttggagaata ttaagaaggc taagttaaa acgcaagttt tctttaaacg tgctgagcaa 180
tacttgatttgc catatcgacg taagaaaag caagagtt 218

```

<210> 70

<211> 293

<212> DNA

<213> *Meloidogyne incognita*

<400> 70

```

taagaaagca gggaaattttt atgtcccaga tgaacctaaa cttgccttttgg tttgtcgat 60
taaggaaatc aacaaggta atttaaattt gcttataaagt ttaggatggg ttttagacaat 120
tcttcctttt taatgttttca taactttttc aaaaaaggta tgattttatc acccattaaat 180
ctacaatttc tttaattttat cagatccatc ctcgtccctcg aaaaggcttca caacttttcc 240
gcttgcgtca aatcaacaat ggagtttca ttaaatttgc taaagctacatc 293

```

<210> 71

<211> 422

<212> DNA

<213> *Meloidogyne incognita*

<400> 71

```

aatgcatttta agactgcttc ggaaggaaag cttaaaggga ttcttgata tactgaggac 60
cagggtttt ctaccgaccc ttctggcgtac actcattcgt ctattttcga cgccgaggcg 120
taagttttga ttttctaaga ttatattaa cttttttaaat ttttcgtct tatgggtctc 180

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AKK110P1									
aaccgcatt	ttgttaaatt	ggtagctgg	tatgataacg	agtttggta	ttcctgccgt	240			
attgttgaact	tgatttagcca	tattgcttcc	aagtctggtt	agatagatgc	ataaaaggga	300			
gaaaagaata	tttttagacga	cattctctaa	aaagtatatt	ttaaatgttag	tttaaatgtat	360			
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<210> 72
<211> 374
<212> DNA
<213> *Meloidogyne incognita*

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<400> 72
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gtggccctcaa gaagagggaaag ctcaggttgc acctactgca Ccaattggtc agccacagcc 180
tcaacagcag caaactcaac aaggaggtga ttggaaactct ggtacttagtg gatggtgaag 240
ggcaggaaaa ttgatagaaaa gagaattat tatggataa atgtaatcaa tggttgttgc 300
tggatattttt gttacatata caacaagttt tattttgttg tttatTTaa aaaaatgtt 360
aattaaaaaaaaaaaa 374

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<210> 73
<211> 120
<212> DNA
<213> *Meloidogyne incognita*

<400> 73
ttttttttttt tttttcttca tcaatattttt gaagtgaaga accagaaga gttgcattcg 60
agctttcaaa ttttgttttt tgattactct tttaaacaaga ttcaactgtat ggatctactg 120

<210> 74
<211> 369
<212> DNA
<213> *Meloidogyne incognita*

<400> 74	gtctaaccaa	tctagagcta	ttcggttcgt	ctgtctgtt	attatttagat	gttgattgaa	60
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tagaattctg	attgtatcg	tcttctttct	cttcctttaa	tggcttttc	aatttatctt	180	
cttccttttc	ttgtccattc	ttttcttcat	tcttttcaaa	aggctcagga	aatttttaatt	240	
cagacccgct	ccttttaact	gctgtatctc	aagaaaaccc	tctaggcaac	gtcccagttc	300	
cactcaatt	caattttgc	aaattttgc	cagatctaag	tccttcttcc	tttgaacgta	360	
attqaactg						369	

<210> 75
<211> 529
<212> DNA
<213> *Meloidoquyne incoqnita*

<400> 75	ttttgttttt	tttttttttt	ttatcagaaaa	aaagtttaat	cagaaaaaaaa	aattaaaaca	60
aatctaaata	aggctctatt	ctaatgttat	atttttcttt	tacataaacc	gtcaaccctc	120	
caagttttc	aatgcttgg	ggttttaatg	gatccctctgg	taataatttg	taggcttagaa	180	
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tataacggtc	agggtcaaaa	ttttggggat	ttgggtatata	ctttggatca	aaaagaacat	360	
ccgatacttg	gggtatcata	aatgtaccc	tagccaacac	aaactttcca	acattcaaata	420	
cttccaaggc	taaatgcccc	aaattgaaag	ggactaaattt	aacgagtctt	aatgtttcat	480	
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<210> 76
<211> 449
<212> DNA
<213> *Meloidoagyne incognita*

AKK110P1

<400> 76

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ga	aggcaca	acgactaaa	caagcaaaac	aggaagcgc	agctgaaatt	gacaaatata	180
ga	gaggaacg	tgaaaaacgt	tttaaagagt	ttgaacatcaa	ttacatcgcc	gctagagatg	240
at	attgctgc	acaataaaag	cgtaaaaactg	atgagacgt	taatgaaatg	actcgtagtg	300
tt	gctgctaa	taaacagcag	gttaattgttc	gtctacttca	acttgcgtgt	gacattcgtc	360
ca	agaactgca	tcacaattha	caacctcaac	ttaagcttaa	tgaaaagcct	gcctaatttg	420
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<210> 77

<211> 643

<212> DNA

<213> Meloidogyne incognita

<400> 77

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ac	gtcccatg	tgcgggcctc	acaagcttcg	tgaatcgctt	cctcttattt	tgttcttcg	180
ta	atctgtcta	aaatatgcac	aatcttataa	tgaagcttag	atgatttgc	aacaacgtct	240
ca	tattaaagtt	gatggcaagg	tgcgtacaga	aatgcgttt	ccagctggat	ttatggatgt	300
gg	tttccatt	gagaaaactg	gcgaagtctt	tcgtcttctc	tatgtatgtc	aaggacgttt	360
cattactcat	cgcatacaaa	aggaagaagg	tca	ttgtgcagg	tagtaaagca	420	
agc	gatttggg	ccaaaacaag	ttccctat	tgttactcat	gatgcccgt	ctattcgct	480
tcc	cgatcc	cacatcaagg	ttgacgacac	tgttgcgtt	gatataaaca	ctggaaaggt	540
ta	acatcac	attagattt	attctgtt	tgttgcgtt	attactggt	gtcacaacat	600
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<210> 78

<211> 584

<212> DNA

<213> Meloidogyne incognita

<400> 78

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aa	ataataaaa	ttgaaatata	ataaaaatga	aatttgcagg	caaaaagagc	aattaattcg	180
ag	atttgcatt	gcctccctaa	cacgttgc	gcaatattca	cgagattggc	aacaatcaca	240
ac	agacaaacaa	aatttcatta	acagtttgg	cccttccccc	cattttatcc	cctcttcagg	300
ca	tatgtatgg	ccccaaacaa	aacaaaaat	atttttggaa	gaaggggaaag	tagaagaacc	360
tt	tagggaa	aatgagaagg	aaaaaagagc	acaaactttt	gttcgtttcg	gaaagagagc	420
ac	aaacacattt	gttcgtttt	gaaaaagggg	acagactttt	gttcgatttt	ggagagattc	480
aa	aaacatcaa	cataacttgt	cagatcaga	gcagttaaaa	actgacaaac	aataaaaatg	540
at	gaatttattt	taaaaatttt	tttaatgtatc	tttaatattaa	aatt		584

<210> 79

<211> 556

<212> DNA

<213> Meloidogyne incognita

<400> 79

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tc	ccgcctga	tcaacagcgt	ttgatcttgc	ctgttgcata	acttgcata	ggacgacact	180
tg	gctgtat	taacatccaa	aaggagtcta	cacttcactt	agtttgcgt	cttcgtgg	240
ga	aaaggttca	cggttcat	gtcgtgtcg	gaaagggtcg	tgctcaact	cctaagg	300
aa	aggcagga	acataagaaa	aagaagcgc	gccgtgttt	ccgtcgccatt	caatataacc	360
gt	cgcttcac	caatgttgc	acttgcgtt	cggttgcgtt	aactccaaac	420	
ct	gcataaga	aatgttgcgt	atcttgcata	atgtatgtt	atataatcaa	tttaatatacat	480
tc	gactntat	gaatgttgcgt	tttatttgcata	tttgcataat	tttgcataat	aaaaaccaag	540
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<210> 80

AKK110P1

<211> 424

<212> DNA

<213> Meloidogyne incognita

<400> 80

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 cgatttggca attcggatg gagtccata tccacctagg cctgcattt ataatgttcc 180
 tccatacctg aatatgttga ctcgaacgtt ttctgtacca aatgtaaatc agtacacggg 240
 tgcaataggt ccttatcgac cagcaaatcc tggttataact tattatagct ataaatgcta 300
 tttccgtat agaaattatc gaggctacac actgacggat gcttactgggt acgaccgtta 360
 ttattatccc tcgccaatat acaaacggtc aatgttccca attagattcc ggcattctga 420
 ctac 424

<210> 81

<211> 89

<212> DNA

<213> Meloidogyne incognita

<400> 81

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 caacanatta cggcccatcc tttgaccctt 89

<210> 82

<211> 168

<212> DNA

<213> Meloidogyne incognita

<400> 82

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 aaagacaaca taatttccaa cttttcaat attatccctt ttaacgggtt gattttgcaa 120
 ctcgctccaa ttcgtcccttc ttcttgatag catatgaatt gctcgAAC 168

<210> 83

<211> 67

<212> DNA

<213> Meloidogyne incognita

<400> 83

aattcatcg ccagacattc agcaattgtt ttgatattac ggaaagaagc ttcacgagac 60
 ccagtac 67

<210> 84

<211> 42

<212> DNA

<213> Meloidogyne incognita

<400> 84

taacacgacg aagaggcgaa acatcaacag cctgacgacg aa

42

<210> 85

<211> 429

<212> DNA

<213> Meloidogyne incognita

<400> 85

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 catcaacttt ttaccattgt tacgtccatg catcatcatc gaacaaacca aacgttcaac 180
 aatcgacaa tgagccccc gaaaacgttt gatttgatcg cgaccacgac tggcggcaa 240
 atatttggcc gatttgcctt taacagcaat ataatccact aaagaagcat cattaaacttc 300
 gatatcgctt aaagaccatt taccaacaa tttatccaa ggaaatcaa ttgttagtcat 360
 ttgcataatcc cttgtccac caggaacatc agttgcggccc caattatcat cagcggtaa 420

AKK110P1

429

accatctcc

<210> 86

<211> 435

<212> DNA

<213> Meloidogyne incognita

<400> 86

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 aattttcttt tcatcatttt ttaatttaaa aaacatttt acaaattaca agaacaacaa 120
 acataattgt ctccctttta ttataaaaatt taaagtttaa taagtttaa aacattctcg 180
 actggagtag gtgtacttag tggtagaa aaggcaaat tagttgttg gtttgaagag 240
 acaaattctt ttgcacaagt agcgagaaga tattcgacag aatttggaaat ggaaccccca 300
 catatggatt tagttaaaaa attacatcaa cttttctca atactggttc tggtagttcaat 360
 ggaataactg aacatttga agttaatcca acaatggaaa catcgacatc ctcaacagag 420
 ggttagcag atccg 435

<210> 87

<211> 501

<212> DNA

<213> Meloidogyne incognita

<400> 87

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 atttgtggcc ctaaagaggg ccgtttgggt ttgggtgtt tacttcagct gccttccacc 180
 aattgttccct tagccaccaa atccgttaaag agtacgtc当地 tggcgtttca acgcatagac 240
 gacgtccatg gctgtgaccg tctttctt ggcgtgtacg caataagttt ccgcgtcgcg 300
 gatcacattt tcaaggaaga ctttcagaac acctcgatc tcctcgtaaa tgagcccgaa 360
 aatacgttt actccacccac gacgtgccaa tcgcccggatt gccgggttgg tgataccctt 420
 gatgatatca cgcaagactt ttcgtggcg cttagcgctt cccttccaa gtccctttcc 480
 gcctttact cgtccggaca t 501

<210> 88

<211> 270

<212> DNA

<213> Meloidogyne incognita

<400> 88

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 agaactagtc tcgagtttt ttttttttt ttttaanaaa ttaacaattt atctcatttt 180
 cctcttccat gaaaatttaac aaaaagacga caacttaatc ccataattaa catcattttt 240
 aagcttcatg cggcatgctt cgaataatgt 270

<210> 89

<211> 286

<212> DNA

<213> Meloidogyne incognita

<400> 89

caagcggttc ccaactcaat gttgttgc当地 tgatactcgta gaacaccagt tctcgccaaac 60
 atagaatagt actcaatctc actgcgtcta aggcttggag tattattcga aataataaca 120
 agtttagctt ttccagaacg aagagtcttc aacgtctcg tggatccaa acaataacttg 180
 cccgattttgg taaccatggc gagacgagca ttgatatttt ctgtggactt tttctgtttt 240
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<210> 90

<211> 391

<212> DNA

<213> Meloidogyne incognita

<400> 90

AKK110P1

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 tccttaggcct gcaattaaca atgttcctcc atacctgaaat atgttactc gaacattttc 180
 tgtaccaat gtaaatcagt acacgggtgc aataggtcct tatcgaccag taaaatcctgt 240
 ctatacttat tatagtata aatgttattt tccgtataga aactatcgag gctacacatt 300
 gacggatgct tattggtacg accgttatta ttattttcg cctatataca aacggtaat 360
 gttccaatt agattccggc actctgacta c 391

<210> 91

<211> 131

<212> DNA

<213> Meloidogyne incognita

<400> 91

attatccaca cacctattgg agctaccctt accaaggaaa atggatgac tatgataatc 60
 caacaaatta ccgcccgttc ttcgaccac gcatcagcgc atcattttca agacctttagt 120
 attacacatc a 131

<210> 92

<211> 571

<212> DNA

<213> Meloidogyne incognita

<400> 92

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 cttcaaaaaa ttcatattt gacgaccagc agcagggtgt tgctgtgtt gttgaccacc 180
 acccccttgc gcttgcaccc ttgttgcgtg tcccttcacg tcaacaggca aattgagttg 240
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 accagcaattt tgattacgca tccgtttgct aggaataaca gcaatttcctt cacaatttcg 480
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 tgcttcttgc acagtttga gagaaccgat t 571

<210> 93

<211> 671

<212> DNA

<213> Meloidogyne incognita

<400> 93

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 aattgtcaag ggtgatgtt ttggaaagga aacgcccattt gttctggtaa tggtggatat 180
 tccctccaat gccgaagtgc tttaaaggagt ggaacttggaa ctttacgtt gtccttggc 240
 gaatcttata gctgtcgagc cagtcacgc tgaagaggca gcttcaaaat acattgatta 300
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 tgcaatttgc caaatagctg ctgcgttgg ggttactgtt ggtatgtga agaaagttat 600
 aatttgggaa aatcattcaa gtacccaaat tccgtatgtt aaacatgtca aagtaattaa 660
 aggtggcacg g 671

<210> 94

<211> 289

<212> DNA

<213> Meloidogyne incognita

<400> 94

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 agctgttattt atcgaaaaac gcaactgtc cagcgcaatg tcggcagcaaa aggccgcgt 120
 tgatcacattt catgattggc actttggaaac aaaagatggc gattgggtttt ctatggccgt 180
 tccttccgat ggttcttattt gaaattccggaa aggtttgatc ttctcatttc caattacaat 240

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tgatgcanaa acgcgtgact ggaaaattgt acaaagatta gaactcgat

289

<210> 95
 <211> 262
 <212> DNA

<213> Meloidogyne incognita

<400> 95

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 acgcgtgctg gttactggag cagctggtca gattggttat tccttggta ttcaaattgc 120
 aaaggagat gtttcggga aagaaaacgc catcgttctg gtaatgttgg atattcctcc 180
 aatggccaa gtgcttaaag gagtggaaact tgaactttac gattgtgcct tggcaaatct 240
 tatactgtc gagccagtca cg 262

<210> 96
 <211> 323
 <212> DNA

<213> Meloidogyne incognita

<400> 96

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 aggatttact tgctgtaat gtaaaaatatt taaaatcgca aggactggct ctagcgaaat 120
 attcaaagcc aactgttaag gttctggttg ttggaaatcc agcagataca aatgctttta 180
 tttgtcaaa atatgcagca gaaaaaattc cgacaaagaa tttcagcgc atgactcgctc 240
 ttgaccataa ccgtgcaatt gcccataatag ctgctcggt tttgggttgc ac 300
 tcaagatagt tataatgtgg gga 323

<210> 97
 <211> 717
 <212> DNA

<213> Meloidogyne incognita

<400> 97

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 ctgttagttat agcaacttgtt ccaccaccac ttccagcacc ctctccatgc atatccaaa 240
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 ttgagctttt ctttgcgttca acaatcagaag acaatcgaag caaataacca tcagttgttt 540
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 ctcgagtaaa agtcatttca tggaaatttgg tcaaaacaaac ttgccttga acctcttcac 660
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<210> 98
 <211> 758
 <212> DNA

<213> Meloidogyne incognita

<400> 98

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 aaaaagaagg atggcttcga tgccaaaaag tttgcattt atttggcttc tgaggaact 180
 gcccgtgcgg ttctcaagac ggctgtggcg cctattgaac gtgtcaagtt gtgtctacag 240
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 gtgatccgtt actttccac gcaagctctc aactttgcgt tcaaggacac ttacaagagg 420
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 tttatgagca aaaatttcc tgggtggat agacctaaca gttgaagagt atcttgcct 540
 ctgtgatacg tataacaacac ttcttcaat tggagattca atgttgcgtt gaggatgtca 600
 tagtaatccctt ctgttacaat cacttaacaa ctcacatcaat tccaatgcctt ctgctcagaa 660
 ttataactcc tcaacaatttgcgaaatgtcgatgtt cgggtttgtt cagatcacca agagaaa 720

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tcaaaaatcga aacagattgt tttaaacgtt tgaaattt 758

<210> 99
 <211> 154
 <212> DNA
 <213> Meloidogyne incognita

<400> 99
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 tgcttattct cgcaggttat tggcaacttca cacatttcta ccaataacaa cgttaccgtt 120
 tataatcaaa ctgttccctca aagttatgcc catt 154

<210> 100
 <211> 125
 <212> DNA
 <213> Meloidogyne incognita

<400> 100
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 aagctcgtga ggaatttgatg cgtatgttgc ctgaagacga acttcgcgt tctgtactcc 120
 tcgtt 125

<210> 101
 <211> 219
 <212> DNA
 <213> Meloidogyne incognita

<400> 101
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 aaatcgtaac tggtatatcc aggctacttg tgccacttca ggagatgggt tttatgaagg 120
 tttggactgg ttgagtaacc aatttgaagaa tcaaggtaaa atgagtctaa ataaaaatgg 180
 agagggggaaa gaggagaggt taattttta aggaaaaaaa 219

<210> 102
 <211> 473
 <212> DNA
 <213> Meloidogyne incognita

<400> 102
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 atggggggaaa aaataggagc aagccaaaaa gccaaaaaaa aattttttt ttaaatgatt 120
 tttgttaaatg tttgttttttcaat ttgtgttcaat ttgttagatgc aatgtcggtt gctttcccttc 180
 cactaaaatt tctctttcct ttcttttctc ttctaaaatt ctttcaaatg cgttccaaacg 240
 aaatttcagc ctcctcttggaa tatttcaact cccaaatacg ctttcaaatgt ttgcctttaa 300
 cgttccaaatg agtaccaat ccagtcatca acttttggaa gtctcccttta ttccaaacggg 360
 cctggatgg aattatcggtt tctgtacttct tcatatcttc atatgaaatg tcgcccagact 420
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<210> 103
 <211> 114
 <212> DNA
 <213> Meloidogyne incognita

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 <211> 255
 <212> DNA
 <213> Meloidogyne incognita

AKK110P1

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 gacctcggtc aatggcgaaa aaaattggaa gggactgtt aagaaattct tggcactgca 180
 caatctgttg ggtgtactgt tcatggacaa catccacatg atattgttga tgcaatccga 240
 agtggaaaaa ttgaa 255

<210> 105
 <211> 571
 <212> DNA
 <213> Meloidogyne incognita

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 tctcccaatt catttaaagt ttcatgtttg tgcggcgcca atgacgacgt ttgcattat 180
 agctatacg actgccagtt ttcatcgaa cccattgcgg cagcggctcg tttgttttag 240
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<210> 106
 <211> 235
 <212> DNA
 <213> Meloidogyne incognita

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 agatagtgtatggacttgata tggctaaaag tattttaaat tgaataaaagg aaaaagaagc 180
 attttaaaga aaatttagatg gaaatgtcga agaaagaaaaaa aaatttattttt tttttt 235

<210> 107
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 <212> DNA
 <213> Meloidogyne incognita

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 aacgtctaaa taatgtataa aatggatata aaaaaaaaaaa aa 702

<210> 108
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 <212> DNA
 <213> Meloidogyne incognita

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 agaaagaaaaa tgccaaagga gatgaagaac ttgttgaaga aaaaaggatca aaaaatataaa 180
 ctcctccatt tgcgtcaca ttttcttca ttattccatt tgcgttgaac tcgtactg 240

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 aattatgagg ttgttgttgt tcctgacgtt tttgattgtc tggagctggg tgaggatcac 420
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<210> 109

<211> 994

<212> DNA

<213> Meloidogyne incognita

<400> 109

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 gtcaaaaacta gccaagccga aattcaagcc tttaaaacgt tcaagagaag agcaaaaaga 180
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 ggaagattt aatgtcgatc aatttgatga tggcggttggaa aatgtcgaa agataataac 420
 gaaattcaga taaaaataac aaagaaaatgt ttataaataaa agctgagttt gccgatatacg 480
 accaaaaat tggatgtttt ttacagaaaa ttggtcaagt tttaaagaaa tatagaagt 540
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<210> 110

<211> 476

<212> DNA

<213> Meloidogyne incognita

<400> 110

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 aaatttgagga gtatcaacgt ttcttcgata tgtttgaccg tggaaagaat ggctatattt 180
 tggccactca aatttgggtt attatgtatg ctatggaaaca agatttgtat gaaaaaaactc 240
 ttccaaaattt aatcccgaaaa ttccgacgcg acggcagcgg caaaatcgaa ttccgacgat 300
 ttccgcctt ggtataactt gtggcagaat ctgttagataa ggacactttt cgaaaaagaat 360
 tgagagaagc ttccgtctc ttccgacaagg agggtaatgg ttacatctct cgtccaaacac 420
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<210> 111

<211> 189

<212> DNA

<213> Meloidogyne incognita

<400> 111

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<210> 112

<211> 164

<212> DNA

<213> Meloidogyne incognita

<400> 112

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 tgacagagat gttcttaacg ttaaatccaa cattgtcttc aggaacagct tcagggagaa 480
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<210> 114
 <211> 314
 <212> DNA
 <213> Meloidogyne incognita

<400> 114
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 cggagaatct tggaaagagta agggaaatgtc cagtttttgt tggtaggtctt ggtgggcttg 180
 gatgtgaaat ttggaaaaat ttggccttat caggatttca aaatattgaa gttattgata 240
 tggacacaat tgaccccttca aatctcaaca gacagttttt gtttcgtgaa cacgtatgtt 300
 gcttatacaa agca 314

<210> 115
 <211> 200
 <212> DNA
 <213> Meloidogyne incognita

<400> 115
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 gacttgcatt ttatggccaa ttttcaattttaa taattttgtgg actagattctt attgatgttc 120
 gaagatggtt aaacgcacca gttgtttctt tggtcgaaatt tgacgaaagaa aacaagccac 180
 ggccaggcac aattatttcca 200

<210> 116
 <211> 471
 <212> DNA
 <213> Meloidogyne incognita

<400> 116
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 gagccattgt tgattgttagg aaaggacaaa ttgtctggaa tggatattcg catccgtgtc 180
 aaagggtgggt gtcatgttgc acaaattttat gcaatttcgac agtcaattgc taaagttttg 240
 gtggcctattt accgaaaaaa cgtggatgag caaagcaaga aagaattgaa ggtatcaactt 300
 gttgcttattt atcgtaattt gcttggcc gatccgagac gtcacgagcc aaagaagttt 360
 ggaggacctg gtgctcgatc tcgttatc aatcttattt gttaaagaaatg atgaaatttt 420
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<210> 117
 <211> 593
 <212> DNA
 <213> Meloidogyne incognita

<400> 117
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ccaaagcatca	tatcattagt	aatgcttcct	gcactactaa	ttgtcttgct	cctttgcga	540
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<211> 576

<212> DNA

<213> Meloidogyne incognita

<400> 118

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<210> 119

<211> 559

<212> DNA

<213> Meloidogyne incognita

<400> 119

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<210> 120

<211> 366

<212> DNA

<213> Meloidogyne incognita

<400> 120

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<211> 661

<212> DNA

<213> Meloidogyne incognita

<400> 121

AKK110P1									
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aaaaaaaaaa	ttttttttat	tttttttcca	taatgtatc	tatatttttt	gctttttaatc	600			
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<210> 122
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<212> DNA
<213> *Meloidogyne incognita*

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<210> 123
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<212> DNA
<213> *Meloidogyne incognita*

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<212> DNA
<213> *Meloidogyne incognita*

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<210> 125
<211> 1013
<212> DNA
<213> *Meloidoqyne incognita*

<400> 125

AKK110P1

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<210> 126

<211> 80

<212> DNA

<213> Meloidogyne incognita

<400> 126

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<210> 127

<211> 585

<212> DNA

<213> Meloidogyne incognita

<400> 127

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cgacatttga	ctctacaatt	gacacacacc	ttttcacaca	tttacaaaat	acattaaaaaa	480
aaaattttttt	ttggcttttt	ggctgtctcc	tattttttcc	ccccatcatt	ctccctattc	540
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<210> 128

<211> 287

<212> DNA

<213> Meloidogyne incognita

<400> 128

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agccaaatcc	aaaactttac	aaaatgcata	tttttgcac	taatcatgtt	gttgctaaat	120
cgcgtttctg	gtactttact	agtagtgc	gtcgatgtt	gaagactaac	ggagagattt	180
tttcgtgtca	ggaggttttt	gaaaagaaga	taggctctgt	aaagaattat	ggaatttggc	240
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<210> 129

<211> 175

<212> DNA

<213> Meloidogyne incognita

<400> 129

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caaataatca	aggttcaacc	gatcaaggct	gccgattgca	aacgtactgg	agttaaacag	120

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ttccacaact cttcaatcaa gtttcctttg ccgcacatcg tgaatgacaa acgtc 175

<210> 130

<211> 599

<212> DNA

<213> Meloidogyne incognita

<400> 130

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 ggagcaatag aacgcttgc tcgcggaggc tcctcagccc tagtaacgtg aaatttctt 180
 gcaatcatcg atttgtgttag tccattttg gctaagacct gttctaagtc ttgttcata 240
 tgtagcataat tgtagcatacg aacatgtgtt cttggtcaca aaggcattgc 300
 tgattggcct ggttagctacg cgagaaatcg gcgggtttat caaactcctc caaacatcca 360
 tctcactgg agtatcccac agggcaggga tttggagggc cacaatatgc tggcaaaaca 420
 ttgtcaactt taatctcttgc gcggtgtgaa aattcagatt ctggatggag ttgttggct 480
 ctttcaccgg cacccctgt cataaattta tgtccaaacg caatggggcc ggaagcactt 540
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<210> 131

<211> 466

<212> DNA

<213> Meloidogyne incognita

<400> 131

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 ttctggacgt tattttcac aaaatttgcgat ggcttagttgc cctaagactg atgtctctt 120
 attggaggat tgcaagaggc ttgggagttac tacagcacat gataatgcac aagttgctcg 180
 tggaaatgtat gtgtgtatggat tagcaggtaa accaactatt gtgtctaaag ttgttgcgg 240
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 acgttacatt gagcagtaat tgacttcaga atcccgaaattt gttcgtgtaa tgccagatac 360
 tcctgttaggt ggttaggagca ggctgtgcgca gccatataatc attggatca gcattgtcag 420
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<210> 132

<211> 266

<212> DNA

<213> Meloidogyne incognita

<400> 132

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 tgcggctttt tgaatatggaa ggtttccgc ctgaagcggaa ttattttttt ttgggtgatt 180
 atgtggatag aggaaagcag agcttggaga cgatttgcgtt gctgtggcc tacaagatca 240
 aatcccccgaa aaattttttt tgctga 266

<210> 133

<211> 308

<212> DNA

<213> Meloidogyne incognita

<400> 133

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 tggaaaacat ttactgtatgg cttcaattgtt ctgcacattt ctgtgtatcg cgtgagaaaa 120
 atattttgtt gccatggagg tttgtcacca gatttgcaga atatggagca aatttgcaga 180
 attatgcac cgacggatgtt gccagataca gttcttctt ggcacccctt atggctgtat 240
 ccagaccaag atgtccaaagg attggggagaa aatgatcgat gggctctttt cacttttggaa 300
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<210> 134

<211> 335

<212> DNA

<213> Meloidogyne incognita

AKK110P1

<400> 134

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tggccgccccg	tgatggaga	agcaggcaaa	attatttaca	agaacattca	attcctcaac	120
tttttgggg	ttaatgact	ggacttata	acaatcaacc	aatcgatcct	attcaatttt	180
tggagaatgc	aatagctaaa	cttcgaaaaa	atcctgatct	tccatcaaag	tgggatactt	240
ttataagtgt	ttcgcctcaa	caacagcaac	aacaacagac	gagaatgaat	actggagaaa	300
atgcagttc	ttataaacaa	agcactccta	tcgaa			335

<210> 135

<211> 506

<212> DNA

<213> Meloidogyne incognita

<400> 135

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ttcctgagac	tttgatcacc	ttcaaaacat	taaaacgaac	agtttactc	aaaggcctgc	180
attaccgat	cgtgacaata	tcaccaatag	agatatcag	gaaacatggc	gaacagtgaa	240
cggacatgtt	tttgtgacgt	ttctcgatc	gacgatattt	cggaacaaaag	tgc当地aaat	300
cacccgaat	gacaatttgg	cgctgcattt	tgttcttgat	aacaacacca	gtcaaaaatac	360
ggcacgaat	tgaacattt	ccagtgaaag	gacattttt	gtcaatataa	ttgccttcga	420
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<210> 136

<211> 230

<212> DNA

<213> Meloidogyne incognita

<400> 136

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ctccagtc	ctacgaaaat	cttgcgag	atcaagggag	taattcgaca	ttatggattc	120
tttttgtgt	tttttaattgt	ttattttgc	tactaatttt	cttcttaattt	gccgccttacc	180
tccgtgtcg	cattttggc	tccgc	ccctt	acaaaaacca	gttccgtcg	230

<210> 137

<211> 216

<212> DNA

<213> Meloidogyne incognita

<400> 137

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tctagtgaat	cggaagaaag	tgatgaacaa	caaagacgg	ggaaatggac	aatctaaca	180
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<210> 138

<211> 395

<212> DNA

<213> Meloidogyne incognita

<400> 138

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gaagacagct	gtatttgtgt	tggcaacact	ccaaacattt	actccagtt	acgggacgg	120
ctctgttctc	gttatgtgtc	acactcgca	acttgctttt	caaatttcaa	agaaatatga	180
aagatttagc	aaatataatgc	ccggaactaa	gttttcgggtt	ttctttgggt	gtatgcccgt	240
caagaaggac	gaggagactt	tggctaagaa	cactccgcac	attgttgg	gcactccagg	300
gcgtctgt	gcgttgggac	gtacaggaca	attgaagctg	aaaacatca	aattcttcgt	360
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<210> 139

<211> 591

AKK110P1

<212> DNA

<213> Meloidogyne incognita

<400> 139

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ctccacacca aactctgggt cattgacaac cggcactttt atctgggttc agcaaacatg 120
gactggcagt cacttactga agtcaaggaa atgggtctta tgctgttgaa ctgctcctgt 180
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gaatttgttc gaatttgcgtt aatggattat attcctgcaa caatttacat gccgaatgg 480
aacaacatata attggccatc gatcgatgac gcgataagaa cggcagctta tcgggggttg 540
aaagttgacc tttgggtgagt ctgtggcccc atttgaatga acgagcgatt t 591